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# COMPOSITIONS COMPRISING XYLANASE AND ENDOGLUCANASE

## Background of the Invention

### Field of the Invention

The present invention relates to compositions comprising at least one xylanase of glycoside hydrolase family 11, and at least one endoglucanase homologous to an endoglucanase produced by a strain of *Thermoascus aurantiacus*, as well as the preparation and use thereof, in particular in relation to animal feed.

### Description of the Related Art

The below references disclose feed additives comprising a xylanase and an endoglucanase, but there is no disclosure of compositions comprising an endoglucanase which is homologous to the *Thermoascus aurantiacus* endoglucanase:

EP 494916 describes feed additives comprising a cellulase and a xylanase producible by means of *Humicola insolens* or *Trichoderma longibrachiatum*;

EP 704167 describes enzyme feed additives comprising a xylanase, a protease and optionally a beta-glucanase;

The product ROXAZYME® G2 marketed by F. Hoffmann-La Roche Ltd. as a feed additive contains an enzyme complex derived from *Trichoderma longibrachiatum*, the main activities of which are cellulase (IUB No. 3.2.1.4), endo-beta-1,3:4-glucanase (IUB No. 3.2.1.6) and xylanase (IUB No. 3.2.1.8).

An endoglucanase derived from *Thermoascus aurantiacus* has been disclosed, but not in a composition together with a family 11 xylanase:

Bhat et al, Spec. Publ. - R. Soc. Chem. (1998), 204-221 describes the purification of an endoglucanase and a xylanase from a strain of *Thermoascus aurantiacus*. However, the xylanase produced by this strain is a family 10 and not a family 11 glycoside hydrolase.

The sequence of an endo-beta-1,4-glucanase from *Thermoascus aurantiacus* IFO 9748 was published as GenPept Accession no. AAL16412 on 23.10.2001.

It is an object of the present invention to provide improved compositions having xylanase and endoglucanase activity.

## Summary of the Invention

The present invention relates to compositions comprising

(i) at least one polypeptide having xylanase activity, the polypeptide being a family 11 glycoside hydrolase; and

(ii) at least one polypeptide having endoglucanase activity, the polypeptide comprising

(a) an amino acid sequence of at least 75 % identity to amino acids 1 to 335, or 31 to 335 of SEQ ID NO:2,

and/or wherein the polypeptide is

(b) encoded by a nucleic acid sequence which hybridizes under low stringency conditions with

(i) the mature endoglucanase encoding part of the plasmid contained in Escherichia coli DSM 14541,

(ii) nucleotides 1 to 1008, or 90 to 1008 of SEQ ID NO:1,

(iii) a subsequence of (i) or (ii) of at least 100 nucleotides, or

(iv) a complementary strand of (i), (ii) or (iii);

(c) a variant of the polypeptide having the amino acid sequence of SEQ ID NO:2 comprising a substitution, deletion, and/or insertion of one or more amino acids,

(d) an allelic variant of (a) or (b); or

(e) a fragment of (a), (b), or (d) that has endoglucanase activity.

The present invention also relates to methods of preparing such compositions, their use in animal feed, their use for treatment of vegetable proteins, as well as animal feed compositions with content thereof.

## Brief Description of the Figures

Figure 1 shows the pH profile of endoglucanase Cel5A derived from *Thermoascus aurantiacus* CGMCC No. 0670;

Figure 2 shows the pH 3 stability of the same;

Figure 3 shows the temperature profile of the same;

Figure 4 shows the temperature stability at 50-70°C of the same;

Figure 5 shows the temperature stability at 85°C of the same;

Figure 6 shows the activity on various substrates of the same;

Figure 7 shows a DNA sequence encoding the same (comprising SEQ ID NO:1);

Figure 8 shows the deduced amino acid sequence of the same (SEQ ID NO:2);

Figure 9 shows a DSC thermogram for the endoglucanase Cel5A derived from *Thermoascus aurantiacus* CGMCC No. 0670; and

Figure 10 shows a DSC thermogram for a xylanase derived from *Thermomyces lanuginosus.*

## Detailed Description of the Invention

### Polypeptides Having Enzyme Activity

The present invention refers to polypeptides having an amino acid sequence which has a certain degree of identity to a specified amino acid sequence, and which have enzymatic activity, e.g. endoglucanase or xylanase activity (hereinafter "homologous polypeptides").

For purposes of the present invention the degree of identity between two amino acid sequences, as well as the degree of identity between two nucleotide sequences, is determined by the program "align" which is a Needleman-Wunsch alignment (i.e. a global alignment). The program is used for alignment of polypeptide, as well as nucleotide sequences. The default scoring matrix BLOSUM50 is used for polypeptide alignments, and the default identity matrix is used for nucleotide alignments. The penalty for the first residue of a gap is -12 for polypeptides and -16 for nucleotides. The penalties for further residues of a gap are -2 for polypeptides, and -4 for nucleotides.

"Align" is part of the FASTA package version v20u6 (see W. R. Pearson and D. J. Lipman (1988), "Improved Tools for Biological Sequence Analysis", PNAS 85:2444-2448, and W. R. Pearson (1990) "Rapid and Sensitive Sequence Comparison with FASTP and FASTA," Methods in Enzymology 183:63- 98). FASTA protein alignments use the Smith-Waterman algorithm with no limitation on gap size (see "Smith-Waterman algorithm", T. F. Smith and M. S. Waterman (1981) J. Mol. Biol. 147:195-197).

In particular embodiments, the polypeptide has the relevant enzymatic activity, and has an amino acid sequence which has a degree of identity to a specified amino acid sequence (a mature polypeptide) of at least about 65%, or of at least about 70%, or of at least about 75% or of at least about 80%, or of at least about 85%, or of at least about 90%, or of at least

about 95%, or of at least about 97%.

In another particular embodiment, these homologous polypeptides have an amino acid sequence which differs by five, four, three, two or only one amino acid(s) from the specified amino acid sequence.

The polypeptides referred to herein may comprise the amino acid sequence specified, or they may be an allelic variant thereof; or a fragment thereof that has the relevant enzyme activity. In one embodiment, the polypeptides comprise the amino acid sequence specified or an allelic variant thereof; or a fragment thereof that has the relevant enzyme activity. In another embodiment, the polypeptides consist of the amino acid sequence specified, or an allelic variant thereof; or a fragment thereof that has the relevant enzyme activity.

A fragment of a specified amino acid sequence is a polypeptide having one or more amino acids deleted from the amino and/or carboxyl terminus of this amino acid sequence. In one embodiment, a fragment contains at least 60 amino acid residues, or at least 68, or at least 70, or at least 75, or at least 100, or at least 150, or at least 160, or at least 170, or at least 180, or at least 190, or at least 200, or at least 210, or at least 220, or at least 240, or at least 260, or at least 280, or at least 300, or at least 310, or at least 320, or at least 330, or at least 334 amino acid residues.

An allelic variant denotes any of two or more alternative forms of a gene occupying the same chromosomal locus. Allelic variation arises naturally through mutation, and may result in polymorphism within populations. Gene mutations can be silent (no change in the encoded polypeptide) or may encode polypeptides having altered amino acid sequences. An allelic variant of a polypeptide is a polypeptide encoded by an allelic variant of a gene.

A mature polypeptide or a mature amino acid sequence refers to that part of an amino acid sequence which remains after a potential signal peptide part has been cleaved off. And analogously, a mature polypeptide encoding part of a gene refers to that part of a gene, which corresponds to a mature polypeptide.

The present invention also refers to polypeptides having a specified enzyme activity and which are encoded by nucleic acid sequences which hybridize under very low stringency conditions, preferably low stringency conditions, more preferably medium stringency conditions, more preferably medium-high stringency conditions, even more preferably high stringency conditions, and most preferably very high stringency conditions with a nucleic acid probe which hybridizes under the same conditions with a specified nucleotide sequence, or a

subsequence or a complementary strand thereof (J. Sambrook, E.F. Fritsch, and T. Maniatus, 1989, *Molecular Cloning, A Laboratory Manual*, 2d edition, Cold Spring Harbor, New York). In one particular embodiment the nucleic acid probe is selected from amongst the specified nucleic acid sequences.

A subsequence may be at least 100 nucleotides, or in another embodiment at least 200 nucleotides. Moreover, the subsequence may encode a polypeptide fragment that has the relevant enzyme activity.

For long probes of at least 100 nucleotides in length, very low to very high stringency conditions are defined as prehybridization and hybridization at 42°C in 5X SSPE, 0.3% SDS, 200 µg/ml sheared and denatured salmon sperm DNA, and either 25% formamide for very low and low stringencies, 35% formamide for medium and medium-high stringencies, or 50% formamide for high and very high stringencies, following standard Southern blotting procedures.

For long probes of at least 100 nucleotides in length, the carrier material is finally washed three times each for 15 minutes using 2 x SSC, 0.2% SDS preferably at least at 45°C (very low stringency), more preferably at least at 50°C (low stringency), more preferably at least at 55°C (medium stringency), more preferably at least at 60°C (medium-high stringency), even more preferably at least at 65°C (high stringency), and most preferably at least at 70°C (very high stringency).

For short probes which are about 15 nucleotides to about 70 nucleotides in length, stringency conditions are defined as prehybridization, hybridization, and washing post-hybridization at 5°C to 10°C below the calculated $T_m$ using the calculation according to Bolton and McCarthy (1962, *Proceedings of the National Academy of Sciences USA* 48:1390) in 0.9 M NaCl, 0.09 M Tris-HCl pH 7.6, 6 mM EDTA, 0.5% NP-40, 1X Denhardt's solution, 1 mM sodium pyrophosphate, 1 mM sodium monobasic phosphate, 0.1 mM ATP, and 0.2 mg of yeast RNA per ml following standard Southern blotting procedures.

For short probes which are about 15 nucleotides to about 70 nucleotides in length, the carrier material is washed once in 6X SCC plus 0.1% SDS for 15 minutes and twice each for 15 minutes using 6X SSC at 5°C to 10°C below the calculated $T_m$.

Still further, the polypeptides referred to herein may be variants of the polypeptides specified comprising a substitution, deletion, and/or insertion of one or more amino acids.

The amino acid sequences of the variant polypeptides may differ from the amino acid

sequence specified by an insertion or deletion of one or more amino acid residues and/or the substitution of one or more amino acid residues by different amino acid residues. Preferably, amino acid changes are of a minor nature, that is conservative amino acid substitutions that do not significantly affect the folding and/or activity of the protein; small deletions, typically of one to about 30 amino acids; small amino- or carboxyl-terminal extensions, such as an amino-terminal methionine residue; a small linker peptide of up to about 20-25 residues; or a small extension that facilitates purification by changing net charge or another function, such as a poly-histidine tract, an antigenic epitope or a binding domain.

Examples of conservative substitutions are within the group of basic amino acids (arginine, lysine and histidine), acidic amino acids (glutamic acid and aspartic acid), polar amino acids (glutamine and asparagine), hydrophobic amino acids (leucine, isoleucine and valine), aromatic amino acids (phenylalanine, tryptophan and tyrosine), and small amino acids (glycine, alanine, serine, threonine and methionine). Amino acid substitutions which do not generally alter the specific activity are known in the art and are described, for example, by H. Neurath and R.L. Hill, 1979, *In*, *The Proteins*, Academic Press, New York. The most commonly occurring exchanges are Ala/Ser, Val/Ile, Asp/Glu, Thr/Ser, Ala/Gly, Ala/Thr, Ser/Asn, Ala/Val, Ser/Gly, Tyr/Phe, Ala/Pro, Lys/Arg, Asp/Asn, Leu/Ile, Leu/Val, Ala/Glu, and Asp/Gly as well as these in reverse.

The polypeptides referred to herein may be encoded by a nucleotide sequence derived from a naturally occurring microorganism, or they may be an analogue, a fragment, a variant, a mutant, or a synthetic polypeptide, which is amended as compared to the one or more wild-type polypeptide(s) on the basis of which it has been designed (genetically engineered). Synthetic or genetically engineered polypeptides, including shuffled enzymes and consensus enzymes, can be prepared as is generally known in the art, eg by Site-directed Mutagenesis, by PCR (using a PCR fragment containing the desired mutation as one of the primers in the PCR reactions), or by Random Mutagenesis. The preparation of consensus proteins is described in eg EP 897985.

The polypeptides referred to herein may be produced or expressed in the original wild-type microbial strain, e.g. in a strain of *Thermoascus aurantiacus*, or in another microbial strain, in a plant, or in an animal - as is generally known in the art. The xylanase and endoglucanase may be co-expressed in one and the same expression host. Also additional enzymes, if any, may be co-expressed.

Accordingly, the polypeptides referred to herein may be wild-type or naturally occurring polypeptides, or they may be genetically engineered or synthetic polypeptides. They may be expressed in the original, wild-type strains or by recombinant gene technology in any other host cell.

Examples of a bacterial polypeptide are a gram positive bacterial polypeptide such as a *Bacillus* polypeptide, or a *Streptomyces* polypeptide; or a gram negative bacterial polypeptide, *e.g.*, an *E. coli* or a *Pseudomonas* sp. polypeptide.

Examples of a *Bacillus* polypeptide are a *Bacillus agaradhaerens, Bacillus circulans, Bacillus licheniformis, Bacillus pumilus,* or *Bacillus subtilis* polypeptide.

Examples of a *Streptomyces* polypeptide are a *Streptomyces coelicolor, Streptomyces lividans, Streptomyces olivaceoviridis, Streptomyces thermocyaneoviolaceus, Streptomyces thermoviolaceus,* or *Streptomyces viridosporus* polypeptide.

Examples of a fungal polypeptide are a yeast polypeptide such as a *Candida, Kluyveromyces, Pichia, Saccharomyces, Schizosaccharomyces,* or *Yarrowia* polypeptide, for example a *Pichia stipitis* polypeptide; or a filamentous fungal polypeptide such as an *Acremonium, Aspergillus, Aureobasidium, Cryptococcus, Emericella, Filibasidium, Fusarium, Gaeumannomyces, Humicola, Lentinula, Magnaporthe, Mucor, Myceliophthora, Neocallimastix, Neurospora, Nocardiopsis, Paecilomyces, Penicillium, Piromyces, Schizophyllum, Talaromyces, Thermoascus, Thermomyces, Thielavia, Tolypocladium,* or *Trichoderma* polypeptide.

In one embodiment, the polypeptide is an *Aspergillus aculeatus, Aspergillus awamori, Aspergillus foetidus, Aspergillus japonicus, Aspergillus kawachii, Aspergillus nidulans, Aspergillus niger, Aspergillus oryzae, Aspergillus tubigensis, Emericella nidulans, Fusarium bactridioides, Fusarium cerealis, Fusarium crookwellense, Fusarium culmorum, Fusarium graminearum, Fusarium graminum, Fusarium heterosporum, Fusarium negundi, Fusarium oxysporum, Fusarium oxysporum f. sp. lycopersici, Fusarium reticulatum, Fusarium roseum, Fusarium sambucinum, Fusarium sarcochroum, Fusarium sporotrichioides, Fusarium sulphureum, Fusarium torulosum, Fusarium trichothecioides, Fusarium venenatum, Gaeumannomyces graminis, Humicola grisea var. thermoidea, Humicola insolens, Humicola lanuginosa, Lentinula edodes, Magnaporthe grisea, Mucor miehei, Myceliophthora thermophila, Neocallimastix frontalis, Neocallimastix patriciarum, Neurospora crassa, Nocardiopsis dassonvillei, Paecilomyces varioti Bainier, Penicillium funiculosum,*

*purpurogenum, Schizophyllum commune, Talaromyces emersonii, Thermoascus aurantiacus, Thermomyces lanuginosus, Trichoderma harzianum, Trichoderma koningii, Trichoderma longibrachiatum, Trichoderma reesei, Trichoderma terrestris,* or *Trichoderma viride* polypeptide.

It will be understood that the definition of the aforementioned species includes both the perfect and imperfect states, and other taxonomic equivalents *e.g.*, anamorphs, regardless of the species name by which they are known. Those skilled in the art will readily recognize the identity of appropriate equivalents.

Strains of these species are readily accessible to the public in a number of culture collections, such as the American Type Culture Collection (ATCC), Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH (DSMZ), Centraalbureau Voor Schimmelcultures (CBS), and Agricultural Research Service Patent Culture Collection, Northern Regional Research Center (NRRL).

Furthermore, such polypeptides may be identified and obtained from other sources including microorganisms isolated from nature (*e.g.*, soil, composts, water, etc.) using the above-mentioned probes. Techniques for isolating microorganisms from natural habitats are well known in the art. The nucleic acid sequence may then be derived by similarly screening a genomic or cDNA library of another microorganism. Once a nucleic acid sequence encoding a polypeptide has been detected with the probe(s), the sequence may be isolated or cloned by utilizing techniques which are known to those of ordinary skill in the art (see, *e.g.*, Sambrook *et al.*, 1989, *supra*).

In a particular embodiment of the composition of the invention, at least one of the component polypeptides is isolated, i.e. essentially free of other polypeptides of enzyme activity, *e.g.*, at least about 20% pure, preferably at least about 40% pure, more preferably about 60% pure, even more preferably about 80% pure, most preferably about 90% pure, and even most preferably about 95% pure, as determined by SDS-PAGE. As it is generally known in the art, for detection purposes the SDS-gel can be stained with Coomassie or silver staining. It should be ensured that overloading has not occurred, e.g. by checking linearity by applying various concentrations in different lanes on the gel.

In another embodiment, at least one of the component polypeptides is well-defined. The term well-defined refers to a preparation of the polypeptide in question which is at least 50% pure as determined by Size-exclusion chromatography. In other particular embodiments

the preparation is at least 60, 70, 80, 85, 88, 90, 92, 94, or at least 95% pure as determined by this method. As it is generally known in the art, following Size-exclusion chromatography, polypeptides can be detected by measuring absorbance at 214 and/or 280 nm.

In still another embodiment, at least one of the component polypeptides is pure, the term pure indicating, that a fractionation of the polypeptide preparation on an appropriate Size-exclusion column reveals only one major polypeptide component having the enzyme activity in question.

The skilled worker will know how to select an appropriate Size-exclusion chromatography column. He might start by fractionating the preparation on e.g. a HiLoad26/60 Superdex75pg column from Amersham Pharmacia Biotech. If the peaks would not be clearly separated he would try different columns (e.g. with an amended column particle size and/or column length), and/or he would amend the sample volume. By simple and common trial-and-error methods he would thereby arrive at a column with a sufficient resolution (clear separation of peaks), on the basis of which the purity calculation can be performed.

In a particular embodiment the endoglucanase polypeptide, as well as the xylanase polypeptide, are isolated and/or well-defined and/or pure. In another embodiment at least one of the additional polypeptides of the composition, if any, is also isolated and/or well-defined and/or pure. In a most preferred embodiment each of the component polypeptides of the composition is isolated and/or well-defined and/or pure.

The use of an isolated and/or well-defined and/or pure polypeptide in the composition of the invention is advantageous. For instance, it is much easier to dose correctly, e.g. to animal feed, enzymes that are essentially free from interfering or contaminating other enzymes. The term dose correctly refers in particular to the objective of obtaining consistent and constant animal feeding results, and the capability of optimising dosage based upon the desired effect.

The composition of the invention can be used for many purposes, for example in animal feed. For such purposes it can be (a) added directly to animal feed (or used directly in a treatment process of vegetable proteins), or (b) it can be used in the production of one or more intermediate compositions such as feed additives or premixes that is subsequently added to feed (or used in a treatment process). The purity indications described above in relation to the terms isolated, well-defined and pure refers to the purity of the component polypeptides,

i.e. before these are mixed to form a composition of the invention, and whether this composition is used according to (a) or (b) above.

Polypeptide preparations with purities of this order of magnitude are in particular obtainable using recombinant methods of production, whereas they are not so easily obtained and also subject to a much higher batch-to-batch variation when the polypeptide is produced by traditional fermentation methods.

**Enzyme Nomenclature - Bernard Henrissat Glycoside Hydrolase Families**

Endoglucanases as well as xylanases are examples of O-Glycoside hydrolase enzymes (EC 3.2.1.x) that hydrolyse the glycosidic bond between two or more carbohydrates or between a carbohydrate and a non-carbohydrate moiety. See the ENZYME site at the internet: http://www.expasy.ch/enzyme/ (ENZYME is a repository of information relative to the nomenclature of enzymes. It is primarily based on the recommendations of the Nomenclature Committee of the International Union of Biochemistry and Molecular Biology (IUB-MB) and it describes each type of characterized enzyme for which an EC (Enzyme Commission) number has been provided (Bairoch A. The ENZYME database, 2000, Nucleic Acids Res 28:304-305). See also the handbook Enzyme Nomenclature from NC-IUBMB, 1992).

The IUB-MB Enzyme nomenclature of glycoside hydrolases is based on their substrate specificity and occasionally on their molecular mechanism; such a classification does not reflect the structural features of these enzymes. A classification of glycoside hydrolases in families based on amino acid sequence similarities has been proposed a few years ago. They currently fall into 87 different families: See the CAZy(ModO) internet site (Coutinho, P.M. & Henrissat, B. (1999) Carbohydrate-Active Enzymes server at URL: http://afmb.cnrs-mrs.fr/~cazy/CAZY/index.html (corresponding papers: Coutinho, P.M. & Henrissat, B. (1999) Carbohydrate-active enzymes: an integrated database approach. In "Recent Advances in Carbohydrate Bioengineering", H.J. Gilbert, G. Davies, B. Henrissat and B. Svensson eds., The Royal Society of Chemistry, Cambridge, pp. 3-12; Coutinho, P.M. & Henrissat, B. (1999) The modular structure of cellulases and other carbohydrate-active enzymes: an integrated database approach. In "Genetics, Biochemistry and Ecology of Cellulose Degradation"., K. Ohmiya, K. Hayashi, K. Sakka, Y. Kobayashi, S. Karita and T. Kimura eds., Uni Publishers Co., Tokyo, pp. 15-23).

**Polypeptides Having Xylanase Activity**

According to the ENZYME site referred to above, xylanases are classified as EC 3.2.1.8. The official name is endo-1,4-beta-xylanase. The systematic name is 1,4-beta-D-xylan xylanohydrolase. Other names may be used, such as endo-(1-4)-beta-xylanase; (1-4)-beta-xylan 4-xylanohydrolase; endo-1,4-xylanase; xylanase; beta-1,4-xylanase; endo-1,4-xylanase; endo-beta-1,4-xylanase; endo-1,4-beta-D-xylanase; 1,4-beta-xylan xylanohydrolase; beta-xylanase; beta-1,4-xylan xylanohydrolase; endo-1,4-beta-xylanase; beta-D-xylanase. The reaction catalysed is the endohydrolysis of 1,4-beta-D-xylosidic linkages in xylans.

According to the CAZy(ModO) site referred to above, xylanases are presently classified in either of the following Glycoside Hydrolyase Families: 10, 11, 43, 5 or 8.

Family 11 glycoside hydrolases can be characterized as follows:

| | |
|---|---|
| CAZy Family: | Glycoside Hydrolase Family 11 |
| Known Activities: | Xylanase (EC 3.2.1.8) |
| Mechanism: | Retaining |
| Catalytic Nucleophile/Base: | Glu (experimental) |
| Catalytic Proton Donor: | Glu (experimental) |
| 3D Structure Status: | Available (see PDB). |
| Fold: | Beta-jelly roll |
| Clan: | GH-C |

The polypeptides having xylanase activity and forming part of the composition of the invention are family 11 glycoside hydrolases, which means they are or can be classified in this family.

In a particular embodiment the family 11 glycoside hydrolase is a fungal xylanase. Fungal xylanases include yeast and filamentous fungal polypeptides as defined above, with the proviso that these polypeptides have xylanase activity.

Examples of fungal xylanases of family 11 glycoside hydrolase are those which can be derived from the following fungal genera: *Aspergillus, Aureobasidium, Emericella, Fusarium, Gaeumannomyces, Humicola, Lentinula, Magnaporthe, Neocallimastix, Nocardiopsis, Orpinomyces, Paecilomyces, Penicillium, Pichia, Schizophyllum, Talaromyces, Thermomyces, Trichoderma.*

Examples of species of these genera are listed above in the general polypeptide section.

The sequences of xylanase polypeptides deriving from a number of these organisms have been submitted to the databases GenBank / GenPept and SwissProt with accession numbers which are apparent from the CAZy(ModO) site.

A preferred fungal xylanase of family 11 glycoside hydrolases is a xylanase derived from

(i) *Aspergillus*, such as SwissProt P48824, SwissProt P33557, SwissProt P55329, SwissProt P55330, SwissProt Q12557, SwissProt Q12550, SwissProt Q12549, SwissProt P55328, SwissProt Q12534, SwissProt P87037, SwissProt P55331, SwissProt Q12568, GenPept BAB20794.1, GenPept CAB69366.1;

(ii) *Trichoderma*, such as SwissProt P48793, SwissProt P36218, SwissProt P36217, GenPept AAG01167.1, GenPept CAB60757.1;

(iii) *Thermomyces* or *Humicola*, such as SwissProt Q43097; or

(iv) a xylanase having an amino acid sequence of at least 75% identity to a (mature) amino acid sequence of a xylanase of (i)-(iii); or

(v) a xylanase encoded by a nucleic acid sequence which hybridizes under low stringency conditions with a mature xylanase encoding part of a gene corresponding to a xylanase of (i)-(iii);

(vi) a variant of the xylanase of (i)-(iii) comprising a substitution, deletion, and/or insertion of one or more amino acids;

(vii) an allelic variant of (i)-(iv);

(viii) a fragment of (i), (ii), (iii), (iv) or (vi) that has xylanase activity; or

(ix) a synthetic polypeptide designed on the basis of (i)-(iii) and having xylanase activity.

Definitions, specific conditions, parameters, as well as particular embodiments of these preferred fungal xylanases forming part of the composition of the invention are apparent from the general polypeptide section hereof (just replace "polypeptide" with "xylanase"). This is for example the case for the calculation of percentage identity, and for the selection of hybridization conditions. This is also the case for particular embodiments relating to, for example, hybridization stringency conditions, or percentage sequence identities.

A preferred xylanase is the *Thermomyces* xylanase of SwissProt Q43097, or analogues thereof as defined in (iv)-(ix) above.

The *Thermomyces* xylanase is also described in WO96/23062. Various *Aspergillus*

xylanases are described in EP 695349, EP 600865, EP 628080, and EP 532533. EP 579672 describes a *Humicola* xylanase.

Xylanase activity can be measured using any assay, in which a substrate is employed, that includes 1,4-beta-D-xylosidic endo-linkages in xylans. Assay-pH and assay-temperature are likewise to be adapted to the xylanase in question. Examples of assay-pH-values are pH 6, 7, 8, 9, 10, or 11. Examples of assay-temperatures are 30, 35, 37, 40, 45, 50, 55, 60, 65, 70 or 80°C.

Different types of substrates are available for the determination of xylanase activity e.g. Xylazyme cross-linked arabinoxylan tablets (from MegaZyme), or insoluble powder dispersions and solutions of azo-dyed arabinoxylan.

For assaying xylanase in feed, premix and the like samples, the enzyme is extracted at temperatures ranging from 50°C up to 70°C (with the higher temperatures used for the more thermostable enzymes) in an extraction media typically consisting of a phosphate buffer (0.1 M and a pH adjusted to the pH optima of the enzyme in question) for a time period of 30 to 60 min.

All measurements are based on spectrophotometric determination principles at approx. 590-600 nm. The enzyme, or the extracted enzyme, as applicable, is incubated with a known amount of substrate and the colour release is measured relative to a standard curve obtained by adding known amounts of an enzyme standard to a similar control diet without enzyme. When no control feed is available, a known amount of enzyme is added to the sample (spiking) and from the differences in response between spiked and non-spiked sample the added amount of enzyme can be calculated.

**Polypeptides Having Endoglucanase Activity**

According to the ENZYME site referred to above, endoglucanases are classified as EC 3.2.1.4. The official name is cellulase. Other names may be used, such as endoglucanase, endo-1,4-beta-glucanase, and carboxymethyl cellulase. The reaction catalysed is endohydrolysis of 1,4-beta-D-glucosidic linkages in cellulose. Also 1,4-linkages in beta-D-glucans also containing 1,3-linkages will be hydrolysed by such enzyme.

According to the CAZy(ModO) site referred to above, endoglucanases are presently classified in either of the following Glycoside Hydrolyase Families: 10, 12, 26, 44, 45, 5, 51, 6, 61, 7, 74, 89, or not yet assigned to a family.

Family 5 glycoside hydrolases can be characterized as follows:

| | |
|---|---|
| CAZy Family: | Glycoside Hydrolase Family 5 |
| Known Activities: | endoglucanase (EC 3.2.1.4); beta-mannanase (EC 3.2.1.78); exo-1,3-glucanase (EC 3.2.1.58); endo-1,6-glucanase (EC 3.2.1.75); xylanase (EC 3.2.1.8); endoglycoceramidase (EC 3.2.1.123) |
| Mechanism: | Retaining |
| Catalytic Nucleophile/Base: | Glu (experimental) |
| Catalytic Proton Donor: | Glu (experimental) |
| 3D Structure Status: | Available (see PDB). |
| Fold: | $(beta/alpha)_8$ |
| Clan: | GH-A |

In a particular embodiment, the polypeptides having endoglucanase activity and forming part of the composition of the present invention are family 5 glycoside hydrolases, which means they are or can be classified in this family.

Examples of family 5 glycoside hydrolases having endoglucanase activity are apparent from the CAZy(ModO) site. Included is, for example, an endoglucanase derived from *Thermoascus aurantiacus* IFO 9748 (GenPept AAL 16412.1).

Applicant is not aware of any other known endoglucanases, be it family 5 glycoside hydrolases or not, which have a percentage of identity to SEQ ID NO:2 of at least 75%.

Definitions, specific conditions, parameters, as well as particular embodiments of the endoglucanases forming part of the composition of the invention are apparent from the general polypeptide section hereof (just replace "polypeptide" with "endoglucanase"). This is for example the case for the calculation of percentage identity, and for the selection of hybridization conditions. This is also the case for particular embodiments relating to, for example, hybridization stringency conditions, percentage sequence identities, or the microorganisms from which the endoglucanase may be derived.

In a particular embodiment, the polypeptide is a polypeptide derived from a filamentous fungus of the phylum *Ascomycota*, preferably of the class *Eurotiomycetidae*,, more preferably of the order *Eurotiales*, even more preferably of the family *Trichocomaceae*.

In another embodiment, the polypeptide is derived from a fungus of the genus *Thermoascus*, for example the species *Thermoascus aurantiacus*, such as the strain

*Thermoascus aurantiacus* CGMCC No. 0670, *e.g.*, a polypeptide with the amino acid sequence of amino acids 1-335, or 31-335 of SEQ ID NO:2.

Endoglucanase activity can be determined using any endoglucanase assay known in the art. For example, various cellulose- or beta-glucan-containing substrates can be applied, under conditions adapted to the enzyme under evaluation (a pH close to the optimum pH and a temperature close to the optimum temperature).

For any of the enzyme types described herein, a preferred assay pH is in the range of 2-10, preferably 3-9, more preferably pH 3 or 4 or 5 or 6 or 7 or 8, for example pH 3 or pH 7. A preferred assay temperature is in the range of 20-80°C, preferably 30-80°C, more preferably 40-75°C, even more preferably 40-60°C, preferably 40 or 45 or 50°C. The enzyme activity is defined by reference to appropriate blinds, e.g. a buffer blind.

An example of an endoglucanase assay using AZCL-Barley beta-Glucan as a substrate is described in Example 3. In a preferred embodiment, this assay is modified to use AZCL-HE-Cellulose as a substrate. In both cases, the degradation of the substrate is followed spectrophotometrically at $OD_{595}$ (see the Megazyme method for AZCL-polysaccharides for the assay of endo-hydrolases (http://www.megazyme.com/booklets/AZCLPOL.pdf).

For the purposes of the present invention, endoglucanase activity is preferably determined according to the procedure described in Example 1, where the enzyme catalyzes the degradation of an Azo-CM-cellulose substrate using a temperature and a pH at which the actual enzyme is active.

In a preferred embodiment of the present invention, the composition comprising endoglucanase and xylanase additionally comprises at least one polypeptide having endo-1,3(4)-beta-glucanase activity, and/or at least one polypeptide having protease activity, and/or at least one polypeptide having phytase activity.

**Polypeptides Having Endo-1,3(4)-beta-glucanase Activity**

According to the ENZYME site referred to above, endo-1,3(4)-beta-glucanases are usually classified as EC 3.2.1.6. The official name is endo-1,3(4)-beta-glucanase. Other names may be used, such as endo-1,4-beta-glucanase, endo-1,3-beta-glucanase, or laminarinase. The reaction catalysed is endohydrolysis of 1,3- or 1,4-linkages in beta-D-glucans when the glucose residue whose reducing group is involved in the linkage to be hydrolysed is itself substituted at C-3. Substrates for this type of enzyme include laminarin,

lichenin and cereal D-glucans.

For the purposes of the present invention, also the following two enzyme classes are included in the term "endo-1,3(4)-beta-glucanase:"

Class EC 3.2.1.73, the official name of which is licheninase. Other names are lichenase, beta-glucanase, endo-beta-1,3-1,4 glucanase, 1,3-1,4-beta-D-glucan 4-glucanohydrolase, or mixed linkage beta-glucanase. The reaction catalysed is hydrolysis of 1,4-beta-D-glycosidic linkages in beta-D-glucans containing 1,3- and 1,4-bonds. This enzyme class acts on lichenin and cereal beta-D-glucans, but not on beta-D- glucans containing only 1,3- or 1,4-bonds.

Class EC 3.2.1.39, the official name of which is glucan endo-1,3-beta-D-glucosidase. Other names are (1-3)-beta-glucan endohydrolase, endo-1,3-beta-glucanase, or laminarinase. The reaction catalysed is hydrolysis of 1,3-beta-D-glucosidic linkages in 1,3-beta-D-glucans. It has a very limited action on mixed-link (1,3-1,4)-beta-D-glucans, but hydrolyses laminarin, paramylon and pachyman.

According to the CAZy(ModO) site referred to above, endo-1,3(4)-beta-glucanases are presently classified in Glycoside Hydrolase Family 16 .

Family 16 glycoside hydrolases can be characterized as follows:

| | |
|---|---|
| CAZy Family: | Glycoside Hydrolase Family 16 |
| Known Activities: | lichenase (EC 3.2.1.73); xyloglucan xyloglucosyltransferase (EC 2.4.1.207); agarase (EC 3.2.1.81); kappa-carrageenase (EC 3.2.1.83); endo-beta-1,3-glucanase (EC 3.2.1.39); endo-beta-1,3-1,4-glucanase (EC 3.2.1.6); endo-beta-galactosidase (EC 3.2.1.103) |
| Mechanism: | Retaining |
| Catalytic Nucleophile/Base: | Glu (experimental) |
| Catalytic Proton Donor: | Glu (experimental) |
| 3D Structure Status: | Available (see PDB). |
| Fold: | Beta-jelly roll |
| Clan: | GH-B |

Examples of endo-1,3(4)-beta-glucanases are apparent from the CAZy(ModO) site.

Endo-1,3(4)-beta-glucanases may be derived as described in the general polypeptide section hereof (just replace "polypeptide" with "endoglucanase").

Endo-1,3(4)-beta-glucanase activity can be determined using any endo-1,3(4)-beta-glucanase assay known in the art. For example, any of the substrates mentioned above can be applied, under conditions adapted to the enzyme under evaluation (e.g. a pH close to the optimum pH and a temperature close to the optimum temperature of the enzyme in question).

A preferred substrate for endo-1,3(4)-beta-glucanase activity measurements is a cross-linked azo-coloured beta-glucan Barley substrate. All measurements are based on spectrophotometric determination principles. For samples of enzyme in feed or premix, the enzyme is extracted at a temperature of 60°C in a 1/30 M Sorensen buffer (0,24 g Dinatriumhydrogenphosphate-Dihydrat (Merck 6580) and 22,47 g Kaliumdihydrogen-phosphate (Merck 4873), in. 4500 ml deionised water, pH is adjusted to 5.00 with HCl and diluted to 50000 ml final volume) following a general procedure similar to that for xylanase determination except that a control feed always must be used to eliminate the endogenous endo-1,3(4)-beta-glucanase background from barley.

Both methods can also be applied to premixes if the premix to be analysed is mixed with a suitable control feed (as described in connection with the assays of Example 1).

For the purposes of the present invention, the endo-1,3(4)-beta-glucanase activity is preferably determined according to the procedure described in Example 1.

For the purposes of the present invention, the polypeptide having endo-1,3(4)-beta-glucanase activity may be the same as, or different from, the polypeptide having endoglucanase activity.

**Polypeptides Having Protease Activity**

The term protease as used herein is an enzyme that hydrolyses peptide bonds (has protease activity). Proteases are also called e.g. peptidases, proteinases, peptide hydrolases, or proteolytic enzymes.

Preferred proteases for use according to the invention are of the endo-type that act internally in polypeptide chains (endopeptidases). Endopeptidases show activity on N- and C-terminally blocked peptide substrates that are relevant for the specificity of the protease in question.

Included in the above definition of protease are any enzymes belonging to the EC 3.4 enzyme group (including each of the thirteen sub-subclasses thereof).

Proteases are classified on the basis of their catalytic mechanism into the following

groupings, each of which is a particular embodiment of a protease for potential use in the composition of the invention: Serine proteases (S), cysteine proteases (C), aspartic proteases (A), metalloproteases (M), and unknown, or as yet unclassified, proteases (U), see Handbook of Proteolytic Enzymes, A.J.Barrett, N.D.Rawlings, J.F.Woessner (eds), Academic Press (1998), in particular the general introduction part.

Protease activity can be measured using any assay, in which a substrate is employed, that includes peptide bonds relevant for the specificity of the protease in question. Assay-pH and assay temperature are likewise to be adapted to the protease in question. Examples of assay-pH-values are pH 5, 6, 7, 8, 9, 10, or 11. Examples of assay temperatures are 25, 30, 35, 37, 40, 45, 50, 55, 60, 65, or 70°C.

Examples of protease substrates are casein, and pNA-substrates, such as Suc-AAPF-pNA (available e.g. from Sigma S-7388). The capital letters in this pNA-substrate refers to the one-letter amino acid code. Another example is Protazyme AK (azurine dyed crosslinked casein prepared as tablets by Megazyme T-PRAK).

Example 2 of WO 01/58276 describes suitable protease assays. A preferred assay is the Protazyme assay of Example 2D (the pH and temperature should be adjusted to the protease in question as generally described previously). For assaying protease in feed or premix, the extraction methods as described herein, e.g. in Example 1 for endoglucanase and xylanase assays, can be used.

In particular embodiments, the protease is a serine protease, a subtilisin protease as defined in WO 01/58275, or a metalloprotease.

Examples of preferred proteases are those described in:

WO 95/02044 (*Aspergillus aculeatus* protease I or protease II);

JP 407 5586 (*Aspergillus niger* acid proteinase (protease A));

Berka et al, Gene 86:153-162, 1993 (*Aspergillus oryzae* aspergillopepsin O);

EP 704167 at p. 8, line 51 to p. 9, line 9;

WO 01/58276 at p. 4, line 25 to p. 5, line 18;

WO 01/58275 at p. 5, line 17 to p. 6, line 5;

the section entitled "Summary of the Invention" of pending patent application DK PA 2001 01821 filed 07.12.01 in the name of Novozymes A/S; or

an analogue, a fragment, a variant, a mutant etc. of any of the above, as described in the general polypeptide part hereof.

**Polypeptides Having Phytase Activity**

In the present context a phytase is an enzyme which catalyzes the hydrolysis of phytate (myo-inositol hexakisphosphate) to (1) myo-inositol and/or (2) mono-, di-, tri-, tetra- and/or penta-phosphates thereof and (3) inorganic phosphate.

According to the ENZYME site referred to above, two different types of phytases are known: A so-called 3-phytase (myo-inositol hexaphosphate 3-phosphohydrolase, EC 3:1.3.8) and a so-called 6-phytase (myo-inositol hexaphosphate 6-phosphohydrolase, EC 3.1.3.26). For the purposes of the present invention, both types are included in the definition of phytase.

For the purposes of the present invention the phytase activity is determined in the unit of FYT, one FYT being the amount of enzyme that liberates 1 micro-mol inorganic ortho-phosphate per min. under the following conditions: pH 5.5; temperature 37°C; substrate: sodium phytate ($C_6H_6O_{24}P_6Na_{12}$) in a concentration of 0.0050 mol/l. Suitable phytase assays are the FYT and FTU assays described in Example 1 of WO 00/20569. FTU is for determining phytase activity in feed and premix. In the alternative, the same principles as described in Example 1, e.g. for endoglucanase and xylanase measurements, can be used for determining phytase activity in feed and premix.

Example of preferred phytases are the following:

*Aspergillus awamori* PHYA (SWISSPROT P34753, Gene 133:55-62 (1993));

*Aspergillus niger (ficuum)* PHYA (SWISSPROT P34752, Gene 127:87-94 (1993));

*Aspergillus awamori* PHYB (SWISSPROT P34755, Gene 133:55-62 (1993));

*Aspergillus niger* PHYB (SWISSPROT P34754, Biochem. Biophys. Res. Commun. 195:53-57(1993));

*Emericella nidulans* PHYB (SWISSPROT O00093, Biochim. Biophys. Acta 1353:217-223 (1997));

*Bacillus subtilis* PHYC_(SWISSPROT O31097, Appl. Environ. Microbiol. 64:2079-2085 (1998));

*Bacillus sp.* PHYT (SWISSPROT O66037, FEMS Microbiol. Lett. 162:185-191 (1998);

*Bacillus subtilis* PHYT_(SWISSPROT P42094, J. Bacteriol. 177:6263-6275 (1995)); and the phytases described in

WO 97/35017 (*Thermomyces lanuginosus* phytase)

WO 98/28408 (*Peniophora* phytase)

WO 98/28409 (Various *Basidiomycete* phytases)

WO 99/49022 (Phytase variants)

WO 99/48380 (Thermostable phytases)

WO 00/43503 (Consensus phytases)

EP 420358 (*Aspergillus niger* phytase)

EP 684313 (Various *Ascomyete* phytases)

EP 0897010 (Modified phytases)

EP 0897985 (Consensus phytases)

US 5830732 (*Schwanniomyces occidentalis* phytase)

US 6110719 (*Escherichia coli* phytase)

AU 724094 (*Bacillus* phytase)

JP 11000164 (*Penicillium* phytase)

WO 97/33976 (*Bacillus* phytase)

US 6139902 (*Aspergillus* phytase)

WO98/13480 (*Monascus anka* phytase)

US patents 5593963, 5770413, 6022846 (transgenic plants expressing phytase)

US patents 5543576, 5714474 (transgenic seeds expressing i.a. phytase); or

an analogue, a fragment, a variant, a mutant etc. of any of the above, as described in the general polypeptide part hereof.

**Thermostable Polypeptides**

In a preferred embodiment, at least one, or at least two, or at least three, or at least four of the polypeptides forming part of the various embodiments of the present invention are thermostable.

In a particular embodiment, the endoglucanase and/or the xylanase and/or the endo-1,3(4)-beta-glucanase, and/or the phytase, and/or the protease are thermostable. In another particular embodiment the xylanase, as well as the endoglucanase and/or the endo-1,3(4)-beta-glucanase thermostable. In still another embodiment, the xylanase, the phytase, and the endoglucanase and/or the endo-1,3(4)-betaglucanase are thermostable.

For the present purposes, the term thermostable means that the polypeptide has a melting temperature, Tm, using Differential Scanning Calorimetry (DSC) of at least 65°C.

In alternative embodiments, the Tm is at least 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 77.5, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99 or 100°C.

For the determination of Tm, an enzyme sample with a purity of at least 90% (or 91, 92, 93, 94, 95, 96, 97, or 98%) as determined by SDS-PAGE may be used. Still further, the enzyme sample may have a concentration of between 0.5 and 2.5 mg/ml protein (or between 0.6 and 2.4, or between 0.7 and 2.2, or between 0.8 and 2.0 mg/ml protein), as determined from absorbance at 280 nm and based on an extinction coefficient calculated from the amino acid sequence of the enzyme in question.

The DSC may take place at pH7 (e.g. in a buffer of 10 mM phosphate, 50 mM NaCl), and with a constant heating rate, e.g. of 1, 1.5, 2, 3, 4, 5, 6, 7, 8, 9 or 10°C/min. Examples of preferred heating rates are 1.0, 1.5 or 2.0°C/min when using an equipment as described in Example 6 herein. For other types of equipment with smaller sample volumes a reliable estimate of Tm can be obtained using a heating rate of, e.g., 3, 4, 5, 6, 7, 8, 9 or 10°C /min, or a heating rate of 20, 30, 40, 50 or even up to 60°C /min.

A thermostable xylanase and a thermostable endoglucanase (also having endo-1,3(4)-beta-glucanase activity), of a Tm of 75.0°C and 77.5°C, respectively, and derived from *Thermomyces lanuginosus* and *Thermoascus aurantiacus,* respectively, are disclosed in Example 6 herein.

Examples of thermostable phytases of a Tm of between 67.0 and 89.3°C are disclosed in WO 99/48380 (see Example 3 thereof).

The present invention also relates to compositions comprising

(i) at least one polypeptide having xylanase activity, and

(ii) at least one polypeptide having endoglucanase activity,

wherein at least one of the polypeptides are thermostable; as well as methods of preparing such compositions, their use in animal feed, their use for treatment of vegetable proteins, and animal feed compositions with content thereof. In a particular embodiment both polypeptides are thermostable. In a further preferred embodiment, at least one of an additional polypeptide of the composition, if any, is also thermostable (eg. an endo-1,3(4)-beta-glucanase, a protease, or a phytase).

**Microorganism Taxonomy**

Questions relating to taxonomy are preferably solved by consulting a taxonomy data base, such as the NCBI Taxonomy Browser which is available at the following internet site: http://www.ncbi.nlm.nih.gov/Taxonomy/taxonomyhome.html/. For questions relating to fungal taxonomy, see preferably Dictionary of the Fungi, 9th edition, edited by Kirk, P.M., P.F. Cannon, J.C. David & J.A. Stalpers, CAB Publishing, 2001.

**Compositions and Uses**

**Animal Feed and Animal Feed Additives**

Xylanase and endoglucanase are essential components of the composition of the invention. These enzymes may be the major enzymatic components of the composition. Also other components may form part of the composition of the invention, for example additional enzymes, and vitamins and/or minerals, examples of which are listed below.

The enzymes of the composition may be prepared in accordance with methods known in the art and may be in the form of liquid enzyme compositions, or dry enzyme compositions such as granulates or microgranulates. The enzymes may be stabilized in accordance with methods known in the art.

A preferred use of the composition of the invention is within the field of animal feed.

For the present purposes, the term animal includes all animals, including human beings. In a particular embodiment, the composition of the invention can be used as a feed additive for non-human animals. Examples of animals are non-ruminants, and ruminants, such as cows, sheep and horses. In a particular embodiment, the animal is a non-ruminant animal. Non-ruminant animals include mono-gastric animals, e.g. pigs or swine (including, but not limited to, piglets, growing pigs, and sows); poultry such as turkeys and chicken (including but not limited to broiler chicks, layers); young calves; and fish (including but not limited to salmon).

The term feed or feed composition means any compound, preparation, mixture, or composition suitable for, or intended for intake by an animal. The feed can be fed to the animal before, after, or simultaneously with the diet. The latter is preferred.

The composition of the invention, when intended for addition to animal feed, may be designated an animal feed additive. Such additive may be a relatively simple mixture of the two enzymes, preferably in the form of stabilized liquid or dry compositions as referred to above. In another type of animal feed additive the two enzymes are in admixture with other

components or ingredients of animal feed. The so-called pre-mixes for animal feed are particular examples of such animal feed additives. Pre-mixes may contain the enzyme(s) in question, and in addition at least one vitamin and/or at least one mineral.

Accordingly, in a particular embodiment, in addition to the component polypeptides, the composition of the invention may comprise or contain at least one fat-soluble vitamin, and/or at least one water-soluble vitamin, and/or at least one trace mineral, and/or at least one macro mineral.

Examples of fat-soluble vitamins are vitamin A, vitamin D3, vitamin E, and vitamin K, e.g. vitamin K3.

Examples of water-soluble vitamins are vitamin B12, biotin and choline, vitamin B1, vitamin B2, vitamin B6, niacin, folic acid and panthothenate, e.g. Ca-D-panthothenate.

Examples of trace minerals are manganese, zinc, iron, copper, iodine, selenium, and cobalt.

Examples of macro minerals are calcium, phosphorus and sodium.

Further, optional, feed-additive ingredients are colouring agents, aroma compounds, and stabilizers.

Additional, optional, enzyme components of the composition of the invention include at least one polypeptide having endo-1,3(4)-beta-glucanase activity, and/or at least one polypeptide having protease activity.

In a particular embodiment, the animal feed additive of the invention is intended for being included (or prescribed as having to be included) in animal diets or feed at levels of 0.0010-12.0%, or 0.0050-11.0%, or 0.0100-10.0%; more particularly 0.05-5.0%; or 0.2-1.0% (% meaning g additive per 100 g feed). This is so in particular for premixes.

Accordingly, the concentrations of the individual components of the animal feed additive, e.g. the premix, can be found by multiplying the final in-feed concentration of the same component by, respectively, 10-10000; 20-2000; or 100-500 (referring to the above three percentage inclusion intervals).

The final in-feed concentrations of important feed components may reflect the nutritional requirements of the animal, which are generally known by the skilled nutritionist, and presented in publications such as the following: NRC, Nutrient requirements in swine, ninth revised edition 1988, subcommittee on swine nutrition, committee on animal nutrition, board of agriculture, national research council. National Academy Press, Washington, D.C.

1988; and NRC, Nutrient requirements of poultry, ninth revised edition 1994, subcommittee on poultry nutrition, committee on animal nutrition, board of agriculture, national research council. National Academy Press, Washington, D.C. 1994.

The xylanase and endoglucanase polypeptides forming part of the composition of the invention should of course be applied in animal feed in an effective amount, i.e. in an amount adequate for improving the nutritional value of the feed. It is at present contemplated that each enzyme is administered in the following dosage ranges: 0.01-200; or 0.01-100; or 0.05-100; or 0.05-50; or 0.10-10 – all these ranges being in mg enzyme protein per kg feed (ppm).

For determining mg xylanase or endoglucanase protein per kg feed, the enzymes are purified from the feed composition, and the specific activity of the purified enzymes is determined using a relevant assay (see under xylanase/endoglucanase activity, substrates, and assays). The xylanase or endoglucanase activity of the feed composition as such is also determined using the same assay, and on the basis of these two determinations, the dosage in mg xylanase or endoglucanase protein per kg feed is calculated.

The same principles apply for determining mg xylanase or endoglucanase protein in feed additives.

Of course, if a sample is available of the xylanase or endoglucanase used for preparing the feed additive or the feed, the specific activity is determined from this sample (no need to purify the enzymes from the feed composition or the additive).

The composition of the invention can be prepared according to methods known in the art, e.g. by mixing the polypeptides having endoglucanase and xylanase activity, and the additional polypeptides, if any.

Animal feed compositions or diets have a relatively high content of protein. An animal feed composition according to the invention has a crude protein content of 50-800, or 75-700, or 100-600, or 110-500, or 120-490 g/kg, and furthermore comprises a composition of the invention.

Furthermore, or in the alternative (to the crude protein content indicated above), the animal feed composition of the invention has a content of metabolisable energy of 10-30, or 11-28, or 11-26, or 12-25 MJ/kg; and/or a content of calcium of 0.1-200, or 0.5-150, or 1-100, or 4-50 g/kg; and/or a content of available phosphorus of 0.1-200, or 0.5-150, or 1-100, or 1-50, or 1-25 g/kg; and/or a content of methionine of 0.1-100, or 0.5-75, or 1-50, or 1-30 g/kg; and/or a content of methionine plus cysteine of 0.1-150, or 0.5-125, or 1-80 g/kg; and/or

a content of lysine of 0.5-50, or 0.5-40, or 1-30 g/kg.

Crude protein is calculated as nitrogen (N) multiplied by a factor 6.25, i.e. Crude protein (g/kg)= N (g/kg) x 6.25 as stated in Animal Nutrition, 4th edition, Chapter 13 (Eds. P. McDonald, R. A. Edwards and J. F. D. Greenhalgh, Longman Scientific and Technical, 1988, ISBN 0-582-40903-9). The nitrogen content can be determined by the Kjeldahl method (A.O.A.C., 1984, Official Methods of Analysis 14th ed., Association of Official Analytical Chemists, Washington DC). But also other methods can be used, such as the so-called Dumas method in which the sample is combusted in oxygen and the amount of nitrous gasses formed are analysed and recalculated as nitrogen.

Metabolisable energy can be calculated on the basis of the NRC publication Nutrient Requirements of Swine (1988) pp. 2-6, and the European Table of Energy Values for Poultry Feed-stuffs, Spelderholt centre for poultry research and extension, 7361 DA Beekbergen, The Netherlands. Grafisch bedrijf Ponsen & looijen bv, Wageningen. ISBN 90-71463-12-5.

In a particular embodiment, the animal feed composition of the invention contains at least one vegetable protein or protein source. Examples of vegetable protein or protein sources are soybean, and the cereals such as barley, maize (corn), oat, rice, rye, sorghum and wheat. Preferred cereals are wheat, barley, oats and rye.

In still further particular embodiments, the animal feed composition of the invention contains 0-80% maize; and/or 0-80% sorghum; and/or 0-70% wheat; and/or 0-70% Barley; and/or 0-30% oats; and/or 0-40% soybean meal; and/or 0-10% fish meal; and/or 0-20% whey.

Animal diets can e.g. be manufactured as mash feed (non-pelleted) or pelleted feed. Typically, the milled feed-stuffs are mixed and sufficient amounts of essential vitamins and minerals are added according to the specifications for the species in question.

The composition of the invention can be added in the form of a solid or liquid enzyme formulation, or in the form of a feed additive, such as a pre-mix. A solid composition is typically added before or during the mixing step; and a liquid composition is typically added after the pelleting step.

The composition of the invention when added to animal feed leads to an improved nutritional value of the feed, e.g. the growth rate and/or the weight gain and/or the feed conversion (i.e. the weight of ingested feed relative to weight gain) of the animal is/are improved. These results may be due to, in turn, one or more of the following effects: Reduction of the viscosity of materials present in the animal's gut; release of nutrients

entrapped e.g. in cell walls of cereals; supplementation and improvement of the endogenous enzyme activities of the animal and the gut microbial flora (this is so in particular in young animals).

In particular embodiments the weight gain is at least 101, 102, 103, 104, 105, 106, 107, 108, 109, or at least 110% of the control (no enzyme addition).

In further particular embodiments the feed conversion is at most (or not more than) 99, 98, 97, 96, 95, 94, 93, 92, 91 or at most 90%, as compared to the control (no enzyme addition).

The composition of the invention may also be used *in vitro*, e.g. to treat vegetable proteins. The term vegetable proteins as used herein refers to any compound, composition, preparation or mixture that includes at least one protein derived from or originating from a vegetable, including modified proteins and protein-derivatives. In particular embodiments, the protein content of the vegetable proteins is at least 10, 20, 30, 40, 50, or 60% (w/w).

Examples of vegetable proteins or protein sources are cereals such as barley, wheat, rye, oat, maize (corn), rice, and sorghum. Other examples are soya bean meal, peas and rape seed meal from leguminosae and brassica families.

The vegetable protein or protein source is typically suspended in a solvent, eg an aqueous solvent such as water, and the pH and temperature values are adjusted paying due regard to the characteristics of the xylanase and endoglucanase in question. The enzymatic reaction is continued until the desired result is achieved, following which it may or may not be stopped by inactivating the enzyme, e.g. by a heat-treatment step.

In another particular embodiment of a treatment process of the invention, the enzyme actions are sustained, meaning e.g. that the enzymes are added to the vegetable proteins or protein sources, but their activity is so to speak not switched on until later when desired, once suitable reaction conditions are established, or once any enzyme inhibitors are inactivated, or whatever other means may have been applied to postpone the action of the enzymes.

## Examples

### Example 1: Enzyme activity assays

**Endoglucanase**

This assay is primarily for assaying endoglucanase activity in animal feed in the form

of mash feed or pellets, or in enzyme premix in powder form. For assaying the endoglucanase activity of enzyme samples which are neither mixed with feed components, nor with vitamins and minerals and the like as in premix, an appropriate starting point is after the heading "incubation and precipitation."

Reagents and solutions

Azo-CM-cellulose solution

Suspend 0.4g Azo-CM-cellulose (Megazyme) in 16ml of demineralised water and stir thoroughly in a boiling water bath for 5 minutes until complete dissolution. After cooling to room temperature 1ml of a 2M sodium acetate buffer, pH4.5 (Megazyme) is added. Adjust the volume with water to 20ml. This solution is kept at 5°C.

Extraction Buffer

Dissolve 5.44g sodium acetate-trihydrate and 6.24g sodium dihydrogen-o-phosphate in approximately 900 ml distilled water and adjust pH to 4.2 with 1N HCl. Add distilled water ad 1000ml.

Precipitation Solution

Dissolve 40g sodium acetate tri-hydrate and 4g zinc acetate in 150ml demineralised water, and adjust to pH5.0 with 5M HCl. Add demineralised water ad 200ml. Add this solution to 800ml ethanol (95%v/v), mix and store at room temperature in a sealed bottle.

Assay Procedure

Pre-treatment of premix

Add 10g premix to 90g corn flour and mix well. Add 10g of this mixture to 90g corn flour and mix well.

Sample preparation and dilution

Weigh 50.0g feed (or premix pre-treated as described above) into a 500ml Erlenmeyer flask and add 500ml Extraction Buffer. Stir for 45 minutes. Take out a sample of 50ml for centrifugation for 10minutes at 2000xg. The supernatants are used for the below enzyme reaction, diluted as required with extraction buffer.

Incubation and precipitation

The incubation temperature is 50°C. 0.1ml of the substrate is pipetted into each vial and pre-incubated for 5 minutes before adding 0.1ml of the supernatant from above. After 60

minutes 0.6ml of the precipitation solution is added to each vial, and the vial is mixed thoroughly on a Vortex mixer. The samples are allowed to stand for 15 minutes at room temperature, and are then mixed again and subjected to centrifugation at 3500 rpm for 10 minutes.

OD measurements and activity calculation

300 microliter of the supernatants from above is immediately pipetted into microtiter plates and the absorbancy at 600nm is measured. The concentration of endoglucanase in the samples is calculated by reference to an appropriate standard curve.

**Xylanase**

This assay is primarily for assaying xylanase activity in animal feed in the form of mash feed or pellets, or in enzyme premix in powder form. For assaying the activity of enzyme samples which are neither mixed with feed components, nor with vitamins and minerals and the like as in premix, an appropriate starting point is after the heading "incubation and precipitation."

Reagents and solutions

Azo-Xylan (Birchwood)

Suspend 0.4 g Azo-Xylan (Birchwood, Megazyme) in 16ml of demineralised water and stir thoroughly in a boiling water bath for 5minutes until complete dissolution. Cool to room temperature and add 1ml 2M sodium acetate buffer, pH 4.5 (Megazyme). Add demineralised water ad 20ml. Store at 5°C.

Extraction Buffer

Dissolve 5.44g sodium acetate-trihydrate and 6.24g sodium dihydrogen-o-phosphate in approximately 900 ml distilled water and adjust pH to 4.2 with 1N HCl. Add distilled water ad 1000ml.

Precipitation Solution

95% (v/v) laboratory grade ethanol is used as the Precipitation Solution.

Assay Procedure

Pre-treatment of premix

Add 10g premix to 90g corn flour and mix well. Add 10g of this mixture to 90g corn

flour and mix well.

Sample preparation and dilution

Weigh 50.0g feed (or premix pre-treated as described above) into a 500ml Erlenmeyer flask and add 500ml Extraction Buffer. Stir for 45 minutes. Take out a sample of 50ml for centrifugation for 10minutes at 2000xg. The supernatants are used for the below enzyme reaction, diluted as required with extraction buffer.

Incubation and precipitation

The incubation temperature is 50°C. 0.125ml of the substrate is pipetted into each vial and pre-incubated for 5 minutes before adding 0.1ml of the supernatant from above. After 150 minutes 0.64ml of the precipitation solution is added to each vial, and the vial is mixed thoroughly on a Vortex mixer. The samples are allowed to stand for 15 minutes at room temperature, and are then mixed again and subjected to centrifugation at 3500 rpm for 10 minutes.

OD measurements and activity calculation

300 microliter of the supernatants from above is immediately pipetted into microtiter plates and the absorbancy at 600nm is measured. The concentration of xylanase in the samples is calculated by reference to an appropriate standard curve.

**Endo-1,3(4)-beta-glucanase**

This assay is primarily for assaying endo-1,3(4)-beta-glucanase activity in animal feed in the form of mash feed or pellets, or in enzyme premix in powder form. For assaying the activity of enzyme samples which are neither mixed with feed components, nor with vitamins and minerals and the like as in premix, an appropriate starting point is after the heading "incubation and precipitation."

Reagents and solutions

Azo-Barley beta-glucan solution

1% Azo-Barley beta-glucan (Megazyme) is used as a substrate.

Extraction Buffer

Dissolve 5.44g sodium acetate-trihydrate and 6.24g sodium dihydrogen-o-phosphate in approximately 900 ml distilled water and adjust pH to 4.2 with 1N HCl. Add distilled water ad 1000ml.

Precipitation Solution

Dissolve 40g sodium acetate tri-hydrate and 4g zinc acetate in 150ml distilled water, and adjust to pH5.0 with concentrated HCl. Add distilled water ad 200ml. Add this solution to 800ml methyl cellosolve (2-methoxyethanol), mix and store at room temperature.

Assay Procedure

Pre-treatment of premix

Add 10g premix to 90g corn flour and mix well. Add 10g of this mixture to 90g corn flour and mix well.

Sample preparation and dilution

Weigh 50.0g feed (or premix pre-treated as described above) into a 500ml Erlenmeyer flask and add 500ml Extraction Buffer. Stir for 45 minutes. Take out a sample of 50ml for centrifugation for 10minutes at 2000xg. The supernatants are used for the below enzyme reaction, diluted as required with extraction buffer.

Incubation and precipitation

The incubation temperature is 50°C. 0.1ml of the substrate is pipetted into each vial and pre-incubated for 5 minutes before adding 0.1ml of the supernatant from above. After 90 minutes 0.5ml of the precipitation solution is added to each vial, and the vial is mixed thoroughly on a Vortex mixer. The samples are allowed to stand for 15 minutes at room temperature, and are then mixed again and subjected to centrifugation at 3500 rpm for 10 minutes.

OD measurements and activity calculation

300 microliter of the supernatants from above is immediately pipetted into microtiter plates and the absorbancy at 600nm is measured. The concentration of endo-1,3(4)-beta-glucanase in the samples is calculated by reference to an appropriate standard curve.

**Specific Enzyme Activity**

For determining specific enzyme activity, the concentration of enzyme protein can be calculated as follows: a) By measuring the absorbance at 280 nm combined with the theoretical molecular weight and the theoretical molar extinction coefficient (both determined from the amino acid sequence); or b) From amino acid analysis. Both methods require a highly purified enzyme sample with full activity

**Examples 2-5**

**Reagents, Media, and Equipment**

Reagents:

Unless otherwise specified, the chemicals used were commercial products of at least reagent grade.

AZCL-substrates from Megazyme:
Azurine-Cross-Linked substrates are supplied as fine powders which are insoluble in buffered solution, but rapidly hydrate to form gel particles which are readily and rapidly hydrolysed by the relevant enzymes, thus releasing the soluble dye-labeled fragment.
AZCL-Barley-beta-Glucan from Megazyme

AZCL-Oat-Spelt-xylan, AZCL-HE-cellulose, AZCL-Potato-Galactan, AZCL-Galactomannan (carob), AZCL-Tamarind-Xyloglucan, AZCL-Debranched-Arabinan

IPTG (Promega, Cat. No. V3951)

X-gal (Promega, Cat. No. V3941)

LMP agarose (Promega, Cat. No. V2111)

Media:
Buffer system (pH 3 to pH 11): 100mM succinic acid, 100mM HEPES, 100mM CHES, 100mM CABS, 1mM $CaCl_2$, 150mM KCl, 0.01% Triton® X-100 adjusted to pH-values 2.0, 2.5, 3.0, 3.5, 4.0, 4.5, 5.0, 6.0, 7.0, 8.0, 9.0, 10.0 or 11.0 with HCl or NaOH (herein for short designated "the succinic acid buffer system").

WB (Wheat Bran medium):
30 g wheat bran, 45 ml of the following solution in each 500 ml shake flask:

- 4 g Yeast Extract
- 1 g $KH_2PO_4$
- 0.5 g $MgSO_4 \cdot 7H_2O$
- 15 g Glucose
- 1000 ml Tap water

Autoclave at 121 °C for 20 min.; pH 5.4 after autoclaving

CBH1 medium:

| | | | |
|---|---|---|---|
| Avicel | 25 g | $(NH_4)_2SO_4$ | 1.4 g |
| $KH_2PO_4$ | 2 g | Urea | 0.3g |
| $CaCl_2.2H_2O$ | 0.3g | $MgSO_4.7H_2O$ | 0.3g |
| $FeSO_4.7H_2O$ | 5mg | $MnSO_4.H_2O$ | 1.6mg |
| Peptone | 1g | Yeast Extract | 10g |
| TWEEN80 | 1ml | Glucose | 5g |
| $H_2O$ | 1000 ml | | |

80 ml in 500 ml Erlenmeyer flask, autoclave 20 minutes under 121°C.

LB liquid medium: To 950ml of deionized $H_2O$, add: 10g bacto-tryptone, 5g bacto-yeast extract, 10g NaCl. Shake until the solutes have dissolved. Adjust the pH to 7.0 with 5N NaOH (~0.2ml). Adjust the volume of the solution to 1 liter with deionized $H_2O$. Sterilize by autoclaving for 20 minutes at 15lb/sq. in. on liquid cycle.

LB plates with ampicillin/IPTG/X-Gal: Add 15g agar to 1 liter of LB medium. Add ampicillin to a final concentration of 100μg/ml, then supplement with 0.5mM IPTG and 80μg/ml X-gal and pour the plates.

1% LMP agarose gel: Add 1g LMP agarose into 100ml 1× TAE buffer.

IPTG stock solution (0.1M):
Add distilled water to 1.2g IPTG to 50ml final volume, filter-sterilize and store at 4°C.

Equipment, including various Kits:
Resource Q column (Amersham Pharmacia, Anion Exchange)
Superdex75 column (Amersham Pharmacia 17-1047-01)
IEF-gel (Amersham Pharmacia 80-1124-80)
Thermomixer comfort (Eppendorf)
RNeasy Mini Kit (QIAGEN, Cat.No.74904)
3' RACE Kit (GIBCO, Cat.No.18373-019) including Adapter primer, and AUAP
dNTP mix (100mM, Promega, Cat. No. U1330)
TaqDNA polymerase system (Promega, Cat. No. M1661) including PCR buffer (200mM Tris-HCl (pH8.4), 500mM KCl)
PCR Preps DNA Purification System (Promega, Cat.No.A7170)
pGEM-T Vector System (Promega, Cat.No.A3600) including T4 DNA Ligase 2XBuffer
JM109 high efficiency competent cells (Promega, Cat. No.L1001)
Minipreps DNA Purification System (Promega, Cat.No.A7100)
BigDye Terminator Cycle Sequencing Ready Reaction Kit (PE Applied Biosystems, Cat. No. 4303149)
ABI Prism 377 DNA sequencer (PE)
5'RACE system (GIBCO, CAT.NO.18374-058) including Abridged Anchor Primer

**Example 2: Cultivation of *Thermoascus aurantiacus* CGMCC No. 0670**

*Thermoascus aurantiacus* CGMCC No. 0670 was grown in WB medium (30g/500 ml flask) at 45°C for 4 days. Enzyme extraction was carried out by adding about 150 ml sterilized water into each shake flask and maintaining at 4°C for at least 4 hours. Supernatant was collected by centrifugation at 7000 rpm for 20 minutes.

**Example 3: Purification of endoglucanase of *Thermoascus aurantiacus* CGMCC No. 0670**

1500 ml supernatant from Example 2 was precipitated with ammonium sulfate (80% saturation) and re-dissolved in 100 ml buffer, ultra-filtrated, and then filtered through a 0.45m filter. The final volume was 30 ml. The solution was applied to a 6 ml Resource Q column equilibrated in 25 mM Tris-HCl buffer, pH 7.4, and the proteins were eluted with a linear NaCl gradient (0 – 0.5M). Fractions from the column were analyzed for endoglucanase activity using the below assay at pH 7.0, and 45°C. Fractions with endoglucanase activity were pooled. Then the pooled solution was ultra-filtrated, and the concentrated solution was applied to a Superdex75 column equilibrated with 25 mM Tris-HCl, pH7.4. The proteins were eluted with the same buffer. Endoglucanase-containing fractions were analyzed by SDS-PAGE and pure fractions were pooled.

Endoglucanase Assay

Substrate: AZCL-beta-Glucan (barley)

Temperature: As desired, e.g. 40, 45, or 50 °C

pH: As desired, e.g. pH 3, or pH 7

Assay buffers (unless otherwise indicated):

200 mM Succinic acid buffer (pH 3)

200 mM Tris-HCl buffer (pH 7)

0.4% AZCL-beta-glucan was suspended in buffer with addition of 0.01% Triton X-100 by gentle stirring. Then a limited amount of this suspension and enzyme samples were mixed in a Microtiter plate or Eppendorf tube and placed on ice before reaction (for amount of substrate and enzyme see the below Results section). The assay was initiated by transferring the Microtiter plate/Eppendorf tube to an Eppendorf thermomixer, which was set to the assay temperature. The plate/tube was incubated for 15-30 minutes on the Eppendorf thermomixer at its shaking rate 700 rpm for Microtiter plate and 1400 rpm for Eppendorf tube reaction. The incubation was stopped by transferring the plate/tube back to the ice bath. Then the tube was centrifuged in an icecold centrifuge for a few minutes and 100/200ml supernatant was transferred to a microtiter plate. $OD_{595}$ was read as a measure of endoglucanase activity. All reactions were done in triplicate and a buffer blind was included in the assay (instead of enzyme).

**Example 4: Characterization of the endoglucanase Cel5A of Thermoascus aurantiacus CGMCC No. 0670**

Three endoglucanases with different profile (pH, temperature, molecular weight, substrate specificity).were purified from culture broth of *Thermoascus aurantiacus* CGMCC No. 0670 grown in WB media.

The one showing endoglucanase activity over relatively wide ranges of pH and temperature was selected for further study.

The purified enzyme was blotted onto a PVDF membrane and N-terminal sequenced. The following sequence was obtained:

N-?LVFTSFGSNESGAEFGSQN.

A homology search showed that this is a family 5 glycoside hydrolase. It is therefore designated endoglucanase Cel5A of *Thermoascus aurantiacus*.

Molecular weight and pI determination of endoglucanase Cel5A

The purity of the purified endoglucanase was verified by SDS-PAGE and IEF gel. The molecular weight of the enzyme is around 32 KDa. Overlay of beta-glucan plate with IEF gel showed that there is only one beta-glucanase activity with pI around 3.5 in the sample.

pH-profile of endoglucanase Cel5A at 45°C

20 ml enzyme sample and 200 ml 0.2% AZCL-beta-glucan in the succinic acid buffer system pH values from pH2.0 to pH11.0 (see above, Media) were mixed in a Microtiter plate and placed on ice before reaction. The assay was initiated by transferring the Microtiter plate to an Eppendorf thermomixer, which was set to the assay temperature 45°C. The plate was incubated for 20 minutes on the Eppendorf thermomixer at 700 rpm shaking rate. The incubation was stopped by transferring the tube back to the ice bath. Then the plate was centrifuged in an ice-cold centrifuge for a few minutes and 100ml supernatant was transferred to a microtiter plate. $OD_{595}$ was read as a measure of beta-glucanase activity. All reactions were done in triplicate and a buffer blind was included in the assay (instead of enzyme).

The resulting pH profile is shown in Fig. 1. In the pH-range of pH 2 to 7, the enzyme retains at least 50% of its maximum activity. The optimum pH is around pH 2.

pH 3 stability of endoglucanase Cel5A at 40°C :

150 ml enzyme sample and 300 ml 0,2M Succinic acid buffer pH 3 were mixed in an Eppendorf tube and incubated under 40°C for 2 hours. Then 100 ml sample was transferred into a new Eppendorf tube with 900 ml 0.4% AZCL-beta-glucan in 0.2M Tris-HCl buffer pH7 with 0.1% Triton X100 and placed on ice before reaction. The assay was initiated by transferring the Eppendorf tube to an Eppendorf thermomixer, which was set to the assay temperature 40°C. The tube was incubated for 30 minutes on the Eppendorf thermomixer at its highest shaking rate (1400 rpm). The incubation was stopped by transferring the tube back to the ice bath. Then the tube was centrifuged in an icecold centrifuge for a few minutes and 200ml supernatant was transferred to a microtiter plate. $OD_{595}$ was read as a measure of endoglucanase activity. All reactions were done in triplicate and a buffer blind was included in the assay (instead of enzyme). For blank, the same amount of substrate, buffer and enzyme were mixed just before reaction start.

The pH 3 stability results are shown in Fig. 2, from which it appears that there is no substantial loss of activity after incubation at pH 3 for 2 hours at 40°C.

Temperature profile of endoglucanase Cel5A at pH 7:

200 ml 0.4% AZCL-beta-glucan in 0.2M Tris-HCl buffer pH 7 and 30 ml enzyme sample were mixed in an Eppendorf tube and put on ice before reaction. The assay was initiated by transferring the Eppendorf tube to an Eppendorf thermomixer, which was set to the assay temperature from 15, 20, 30, 40, 50, 60, 70, 80°C. The tube was incubated for 30 minutes on the Eppendorf thermomixer at its highest shaking rate (1400 rpm). The incubation was stopped by transferring the tube back to the ice bath. Then the tube was centrifuged in an icecold centrifuge for a few minutes and 200ml supernatant was transferred to a microtiter plate. $OD_{595}$ was read as a measure of endoglucanase activity. All reactions were done in triplicate and a buffer blind was included in the assay (instead of enzyme).

The results are shown in Fig. 3, from which it appears that the enzyme is active within the whole temperature range of 20 to 80°C. The optimum temperature is around 70°C. At 40°C and 80°C, the relative activity is 58% and 37%, respectively (relative to the activity at 70°C).

Thermostability at 50, 60, 70 and 85°C of endoglucanase Cel5A at pH 7.4:

100 ml enzyme sample (pH7.4) in an Eppendorf tube was incubated for 10 and 20

minutes on the Eppendorf Thermomixer at 50, 60, 70°C and 300 rpm shaking. For stability at 85°C, the same method was applied but with sampling time as 0, 2, 5 and 10 minutes. The incubation was stopped by transferring the tube back to the ice bath. Un-incubated sample was used as control. The 30 ml of the above incubated sample was transferred into a new Microtiter plate and 200 ml 0.4% AZCL-beta-glucan in 0.2M Tris-HCl buffer pH 7 was added. The assay was initiated by transferring the Microtiter plate to an Eppendorf thermomixer, which was set to the assay temperature 40°C. The plate was incubated for 30 minutes on the Eppendorf thermomixer at 700rpm shaking rate. The incubation was stopped by transferring the tube back to the ice bath. Then the plate was centrifuged in an icecold centrifuge for a few minutes and 100 ml supernatant was transferred to a microtiter plate. $OD_{595}$ was read as a measure of endoglucanase activity. All reactions were done in triplicate and a buffer blind was included in the assay (instead of enzyme).

The results are shown in Fig. 4 (50-70°C), and in Fig. 5 (85°C). The enzyme appears to fully retain its activity after having been incubated for 10 to 20 minutes at a temperature in the range of 50 to 70°C. Also after incubation at 85°C for 10 minutes, the enzyme seems to fully retain its activity.

Substrate specificity of endoglucanase Cel5A at pH 3 and 50°C on various cellulase and hemicellulase substrates:

400 ml 0.2% AZCL-substrate (xylan, HE-cellulose, Galactan, Mannan, Xyloglucan, Arabinan) in 0.2M succinic acid buffer pH 3 with 0.01% Triton X100 and 30 ml enzyme sample (5 x dilution by 0.2 M succinic acid buffer) were mixed in an Eppendorf tube and put on ice before reaction. The assay was initiated by transferring the Eppendorf tube to an Eppendorf thermomixer, which was set to the assay temperature 50°C. The tube was incubated for 15 minutes on the Eppendorf thermomixer at its highest shaking rate (1400 rpm). The incubation was stopped by transferring the tube back to the ice bath. Then the tube was centrifuged in an icecold centrifuge for a few minutes and 200 ml supernatant was transferred to a microtiter plate. $OD_{595}$ was read as a measure of endoglucanase activity. All reactions were done in triplicate and a buffer blind was included in the assay (instead of enzyme).

From the results which are shown in Fig. 6, it appears that the enzyme can degrade beta-glucan and HE-Cellulose, but it has no or very low activity on xylan, arabinan, mannan,

xyloglucan.

**Example 5: Cloning of the gene encoding endoglucanase Cel5A of *Thermoascus aurantiacus* CGMCC 0670**

The gene fragment encoding endoglucanase Cel5A was cloned by RT-PCR from *Thermoascus aurantiacus* CGMCC 0670 as described below.

Sequence analysis of the cDNA clone showed that the sequence contains a coding region of 1005 nucleotides. The coding region including C-terminal extension is shown in Fig. 7. The start and stop codons, ATG and TGA, respectively, are boxed. SEQ ID NO:1 represents the coding part of the Fig. 7 sequence, starting with ATG and ending with TGA.

The translation product of the coding region is shown in Fig. 8. This protein is 335 amino acids in length. Expectedly, amino acid residues 1 to 30 constitute a signal-peptide part, and amino acid residues 31 to 335 constitutes the catalytic domain.

Cultivation and isolation of mycelium

*Thermoascus aurantiacus* CGMCC 0670 was grown in CBH1 medium at 45°C and 165 rpm for 3 days. Then the mycelium was harvested by centrifugation at 7000 rpm for 30 minutes. Harvested mycelium was stored at minus 80°C before being used for extraction of RNA.

Extraction of total RNA

The total RNA was extracted from 100 mg of the mycelium isolated above using the RNeasy Mini Kit.

Design of degenerate primers

Degenerate primers were designed based on determined N-terminal amino acid sequence N-?LVFTSFGSNESGAEFGSQN.

1 5' AA(T/C) GA(A/G) TC(T/C/A/G) GG(T/C/A/G) GC(T/C/A/G) GAA TT 3'

2 5' AA(T/C) GA(A/G) TC(T/C/A/G) GG(T/C/A/G) GC(T/C/A/G) GAG TT 3'

3 5' AA(T/C) GA(A/G) AG(T/C) GG(T/C/A/G) GC(T/C/A/G) GAA TT 3'

4 5' AA(T/C) GA(A/G) AG(T/C) GG(T/C/A/G) GC(T/C/A/G) GAG TT 3'

Cloning of the 3' end of the endoglucanase

The 3' RACE kit was used to synthesize the cDNA of the endoglucanase. About 5mg total RNA was used as template and the Adapter Primer (provided by the 3'RACE system) was used to synthesize the first strand of cDNA. Then the cDNA was amplified by using different degenerate primers. The PCR reaction system and conditions were as follows:

| | |
|---|---|
| 10xPCR buffer | 5μl |
| 25mM $MgCl_2$ | 3μl |
| 10mM dNTP mix | 1μl |
| 3'Primer (10μM) | 1μl |
| AUAP (10μM, provided by 3'RACE system) | 1μl |
| TaqDNA polymerase (5u/μl, Promega) | 0.5μl |
| cDNA synthesis reaction | 2μl |
| Add autoclaved, distilled water to | 50μl |

Conditions:

| | | |
|---|---|---|
| 94°C | 3min | |
| 94°C | 40sec | |
| 55°C | 40sec | 30 cycles |
| 72°C | 1min | |
| 72°C | 10min | |

Gel analysis of the PCR product gave a specific band about ~1kb fragment using primer 2 and primer 3, and the products were recovered from 1% LMP agarose gel, and purified by incubation at 70°C followed by using PCR Preps DNA Purification System. The concentration of the purified products was determined by measuring the absorbances $A_{260}$ and $A_{280}$ in a spectrophotometer. Then these purified fragments were ligated to pGEM-T Vector (Promega kit, Cat.No.A3600):

| | |
|---|---|
| T4 DNA Ligase 2 x Buffer5μl | |
| pGEM-T Vector (50ng) | 1μl |
| PCR product | 50ng |
| T4 DNA Ligase (3 Weiss units/μl) | 1μl |

| | |
|---|---|
| $dH_2O$ to a final volume of | 10μl |

Conditions:

Incubate the reactions overnights at 4°C.

Then we transformed 2-4μl ligation products into 50μl JM109 high efficiency competent cells by the "heat shock" method (J. Sambrook, E.F.Fritsch, T.Maniatis (1989) Molecular Cloning 1.74, 1.84). Transformation cultures were plated onto the LB plates with ampicillin/IPTG/X-Gal, and these plates were incubated overnight at 37°C. Recombinant clones were identified by colour screening on indicator plates and colony PCR screening. The positive clones were inoculated into 3ml LB liquid medium and incubated overnight at 37°C with shaking (~250rpm). The cells were sedimented by centrifugation for 5min at 10,000xg, and a plasmid sample was prepared from the cell pellet by using Minipreps DNA Purification System. Finally, the plasmids were sequenced with BigDye Terminator Cycle Sequencing Ready Reaction Kit by using ABI377 sequencer. The sequencing reaction was as follows:

| | |
|---|---|
| Terminator Ready Reaction Mix | 8μl |
| Plasmid DNA | 1-1.5μg |
| Primer | 3.2pmol |
| $dH_2O$ to a final volume of | 20 μl |

The sequencing result showed that the PCR band obtained using primer 2, as well as primer 3 corresponds to the 3'end of the endoglucanase encoding sequence.

Cloning of 5' end of the endoglucanase

Based on the 3'-end sequence, we designed four specific primers which were used for 5' end sequence cloning.

| | | | |
|---|---|---|---|
| 5'-1 | 5' | AAG ATG TAC TGG GAA GTG | 3' |
| 5'-2 | 5' | TGG TTG AGA TTG AGG ACT AAG | 3' |
| 5'-3 | 5' | GAT TAT AGA ATT GTA GTA TCT | 3' |
| 5'-4 | 5' | AGA GCC GGT CAT TGA GTT G | 3' |

The 5'RACE system was used to synthesize the 5'end fragment of the endoglucanase. 5 mg total RNA and primer 5'-1 was added for synthesis of the first strand. Then other primers were used for the second strand synthesis. The system and conditions of PCR of dC-tailed cDNA is as following:

| | |
|---|---|
| 10xPCR buffer (200mMTris-HCl(pH8.4), 500mM KCl) | 5µl |
| 25mM $MgCl_2$ | 3µl |
| 10mM dNTP mix | 1µl |
| 5'Primer (10µM) | 2µl |
| Abridged Anchor Primer(10µM, provided by 3'RACE system) | 2µl |
| TaqDNA polymerase (5u/µl) | 0.5µl |
| dC-tailed cDNA | 5µl |
| Add autoclaved, distilled water to | 50µl |

PCR Conditions:

| | | |
|---|---|---|
| 94°C | 2min | |
| 94°C | 40sec | |
| 53°C | 40sec | 30 cycles |
| 72°C | 1min | |
| 72°C | 10min | |

Two specific bands corresponding to approx. 700bp and 400 bp resulted from using primers 5'-2, and 5'-4, respectively, using the 5'RACE system. The PCR-products were purified, ligated into the pGEM-T-vector, transformed into JM109 competent cells, and sequenced. The sequencing result showed we got the 5'end fragment of BG025.

Cloning of the full length endoglucanase gene

According to the above 3' and 5' end sequences, two primers for full length cloning were designed:

| | | | |
|---|---|---|---|
| CDS-1 | 5' | ATG AAG CTC GGC TCT CTC GT | 3' |
| CDS-2 | 5' | CTT GTC TCC TGT CTC GTT CAC | 3' |

Primer CDS-1 and AUAP was used for amplifying the full length gene from the cDNA. The following PCR reaction system and conditions were used:

| | |
|---|---|
| 10xPCR buffer | 5µl |
| 25mM $MgCl_2$ | 3µl |
| 10mM dNTP mix | 1µl |
| Primer CDS-1 (10µM) | 1µl |
| AUAP (10µM) | 1µl |
| TaqDNA polymerase (5u/µl) | 0.5µl |
| cDNA synthesis reaction | 2µl |
| Add autoclaved, distilled water to | 50µl |

Conditions:

| | | |
|---|---|---|
| 95°C | 2min | |
| 95°C | 40sec | |
| 58°C | 40sec | 30 cycles |
| 72°C | 1.5min | |
| 72°C | 10min | |

From this amplification, a specific band with the size of about 1.2kb was obtained and this was recovered from gel with the PCR Preps DNA Purification System. Then the purified fragment was ligated into the pGEM-T Vector, and transformed into the competent cells (JM109). Positive clones were screened by colony PCR, and the plasmid was extracted from these clones with Minipreps DNA Purification System. Finally the plasmid was sequenced with the BigDye Terminator Cycle Sequencing Ready Reaction Kit, and the full length endoglucanase encoding sequence was obtained.

**Example 6: Determination of the Thermostability of Enzymes by Differential Scanning Calorimetry (DSC)**

Endoglucanase

The purity of the purified endoglucanase resulting from Example 3 was determined by SDS-PAGE to be above 90%. The concentration of protein was determined to 1.9 mg/ml

(based on $OD_{280}$ and an extinction coefficient calculated on the basis of the amino acid sequence).

For determining the denaturation or melting temperature of the endoglucanase (Td or Tm, respectively), the sample is dialysed over-night at 4°C against a buffer containing 10mM sodium phosphate, 50mM sodium chloride, pH7.0. The dialysed sample was measured against pure buffer in a Microcalorimeter (VP-DSC from Microcal) from 20°C to 95-100°C with a temperature gradient of 1.5°C/min. The melting temperature was determined as the summit of the peak in the resulting thermogram (see Figure 9): Tm 77.5 °C.

Xylanase

A sample of the xylanase derived from *Thermomyces lanuginosus* (see Examples 1-3 of WO 96/23062) of a purity of above 90% as determined by SDS-PAGE, and a concentration of protein of 0.8 mg/ml (based on $OD_{280}$ and an extinction coefficient calculated on the basis of the amino acid sequence) was subjected to a procedure as described above, and Tm was determined to 75.0°C.

**Deposit of Biological Material**

The following biological material has been deposited under the terms of the Budapest Treaty with DSMZ (DSMZ-Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH, Mascheroder Weg 1b, D-38124 Braunschweig, Germany), and CGMCC (the China General Microbiological Culture Collection Center, Institute of Microbiology, Chinese Academy of Sciences, Haidian, Beijing 100080, China); and given the following accession numbers:

| Deposit | Accession Number | Date of Deposit |
|---|---|---|
| *Escherichia coli* | DSM 14541 | 2001-09-28 |
| *Thermoascus aurantiacus* | CGMCC No. 0670 | 2001-12-27 |

The *Escherichia* coli strain harbours a plasmid containing the nucleic acid sequence of endoglucanase Cel5A of *Thermoascus aurantiacus* DSM 14541 (i.e. SEQ ID NO: 1 encoding SEQ ID NO:2).

These strains have been deposited under conditions that assure that access to the cultures will be available during the pendency of this patent application to one determined by

the Commissioner of Patents and Trademarks to be entitled thereto under 37 C.F.R. §1.14 and 35 U.S.C. §122. The deposits represent a substantially pure culture of the deposited strains. The deposits are available as required by foreign patent laws in countries wherein counterparts of the subject application, or its progeny are filed. However, it should be understood that the availability of a deposit does not constitute a license to practice the subject invention in derogation of patent rights granted by governmental action.

*Thermoascus aurantiacus* strain no. CGMCC 0670 was isolated from a soil sample collected on July 21, 1998 in the Yunnan Province, Xishuangbanna, China.

The invention described and claimed herein is not to be limited in scope by the specific embodiments herein disclosed, since these embodiments are intended as illustrations of several aspects of the invention. Any equivalent embodiments are intended to be within the scope of this invention. Indeed, various modifications of the invention in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description. Such modifications are also intended to fall within the scope of the appended claims. In the case of conflict, the present disclosure including definitions will control.

Various references are cited herein, the disclosures of which are incorporated by reference in their entireties.

## Claims

1. A composition comprising

(i) at least one polypeptide having xylanase activity, the polypeptide being a family 11 glycoside hydrolase; and

(ii) at least one polypeptide having endoglucanase activity, the polypeptide comprising

(a) an amino acid sequence of at least 75 % identity to amino acids 1 to 335, or 31 to 335 of SEQ ID NO:2, and/or wherein the polypeptide is

(b) encoded by a nucleic acid sequence which hybridizes under low stringency conditions with

(i) the mature endoglucanase encoding part of the plasmid contained in Escherichia coli DSM 14541,

(ii) nucleotides 1 to 1008, or 90 to 1008 of SEQ ID NO:1,

(iii) a subsequence of (i) or (ii) of at least 100 nucleotides, or

(iv) a complementary strand of (i), (ii) or (iii);

(c) a variant of the polypeptide having the amino acid sequence of SEQ ID NO:2 comprising a substitution, deletion, and/or insertion of one or more amino acids,

(d) an allelic variant of (a) or (b), or

(e) a fragment of (a), (b), or (d) that has endoglucanase activity.

2. The composition of claim 1, wherein the polypeptide having endoglucanase activity is a family 5 glycoside hydrolase.

3. The composition of any one of claims 1-2, wherein at least one of the polypeptides having endoglucanase or xylanase activity is thermostable.

4. The composition of any one of claims 1-3, wherein the polypeptide having xylanase activity is derived from a strain of *Aspergillus*, *Humicola*, *Thermomyces*, or *Trichoderma*.

5. The composition of any one of claims 1-4, which further comprises at least one polypeptide having endo-1,3(4)-beta-glucanase activity, and/or at least one polypeptide having protease activity, and/or at least one polypeptide having phytase activity.

6. The composition of claim 5, wherein at least one of the further polypeptides is thermostable.

7. The composition of any one of claims 1-6, further comprising

(a) at least one fat soluble vitamin, and/or

(b) at least one water soluble vitamin, and/or

(c) at least one trace mineral, and/or

(d) at least one macro mineral.

8. The composition of any one of claims 1-7 which is an animal feed additive.

9. A method of preparing a composition of any one of claims 1-8, the method comprising the step of mixing the polypeptides having endoglucanse and xylanase activity.

10. Use of the composition of any one of claims 1-8 in animal feed.

11. Use of the composition of any one of claims 1-8 in the preparation of animal feed.

12. A method for improving the nutritional value of an animal feed, wherein the composition of any one of claims 1-8 is added to the feed.

13. An animal feed composition having a crude protein content of 50 to 800 g/kg and comprising the composition of any one of claims 1-8.

14. The animal feed of claim 13, which comprises at least one of wheat, barley, oats or rye.

15. A method for the treatment of vegetable proteins, comprising the step of adding the composition of any one of claims 1-8 to at least one vegetable protein or protein source.

16. The method of claim 15, wherein wheat, barley, oats and/or rye is included amongst the at least one vegetable protein source.

**Abstract**

The present invention relates to a composition comprising a xylanase of glycoside hydrolase family 11, and an endoglucanase which is homologous to a thermostable glycoside hydrolase family 5 endoglucanase derived from *Thermoascus aurantiacus*. Preferred xylanases are derived from *Aspergillus, Humicola, Thermomyces* and *Trichoderma*. The composition is particularly useful for animal feed purposes. Optional additional components are vitamins, minerals, or additional enzymes such as proteases, and/or endo-1,3(4)-beta-endoglucanases, and/or phytases. One or more of the enzymes are preferably thermostable.

**Fig. 1**

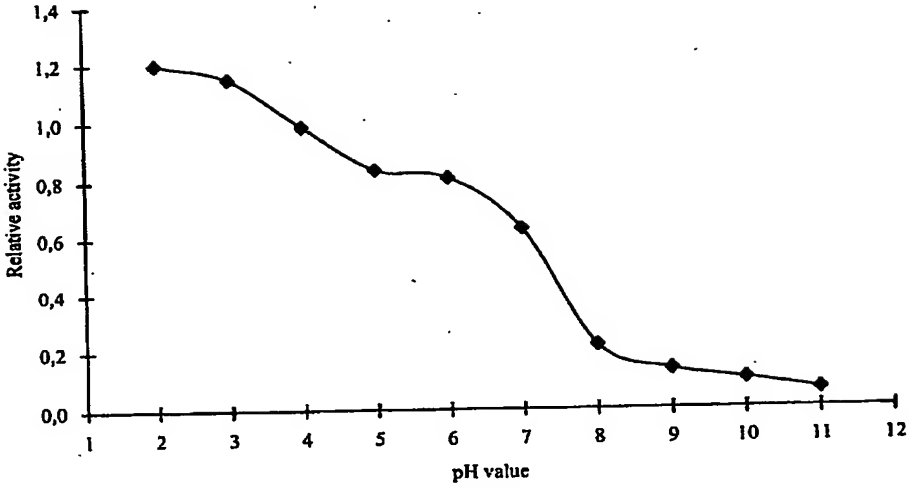


**Fig. 2**

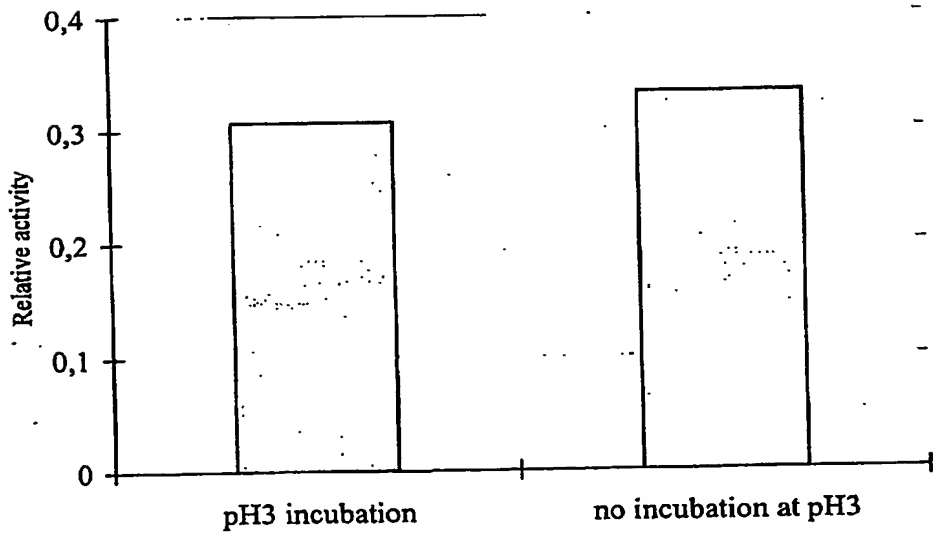

Fig. 3

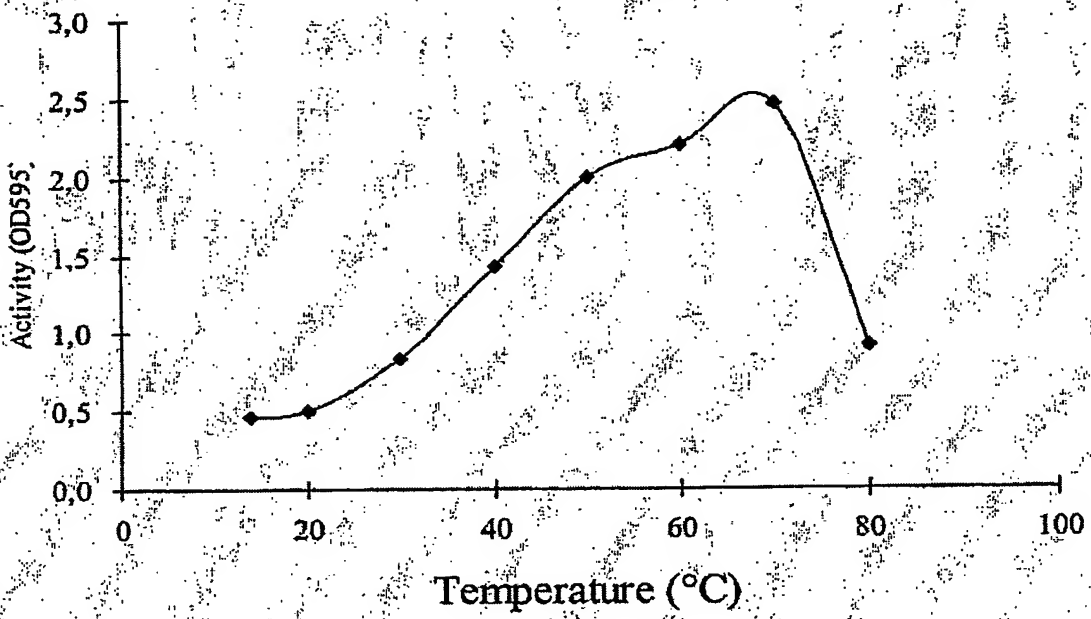


Fig. 4

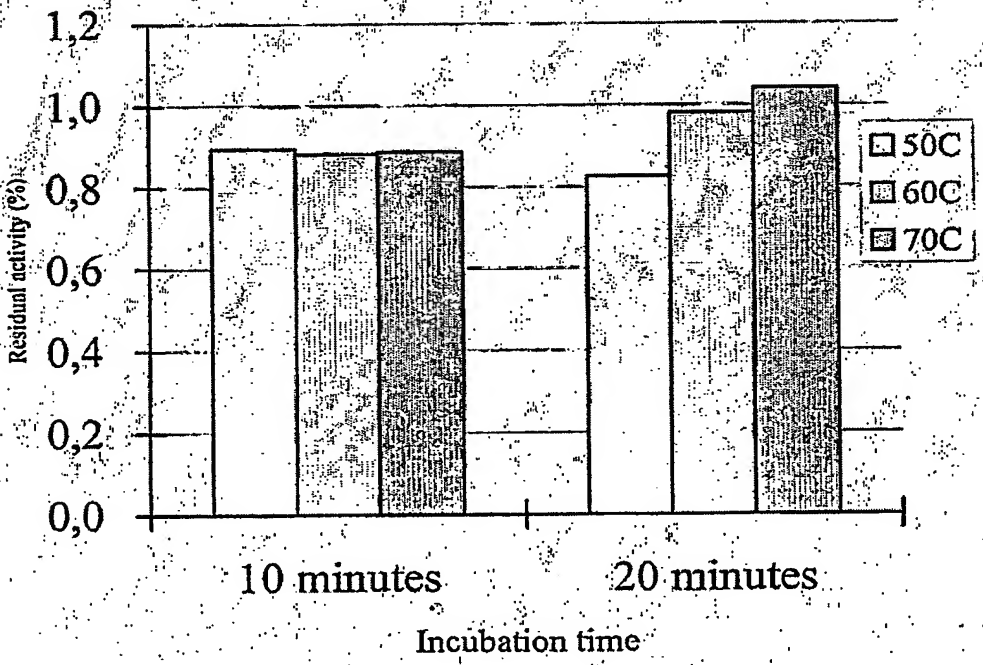

Fig. 5

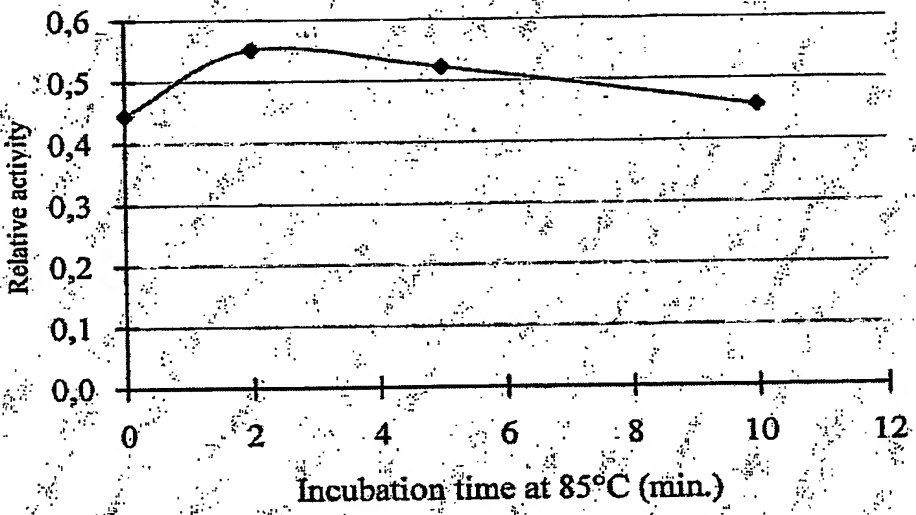


Fig. 6

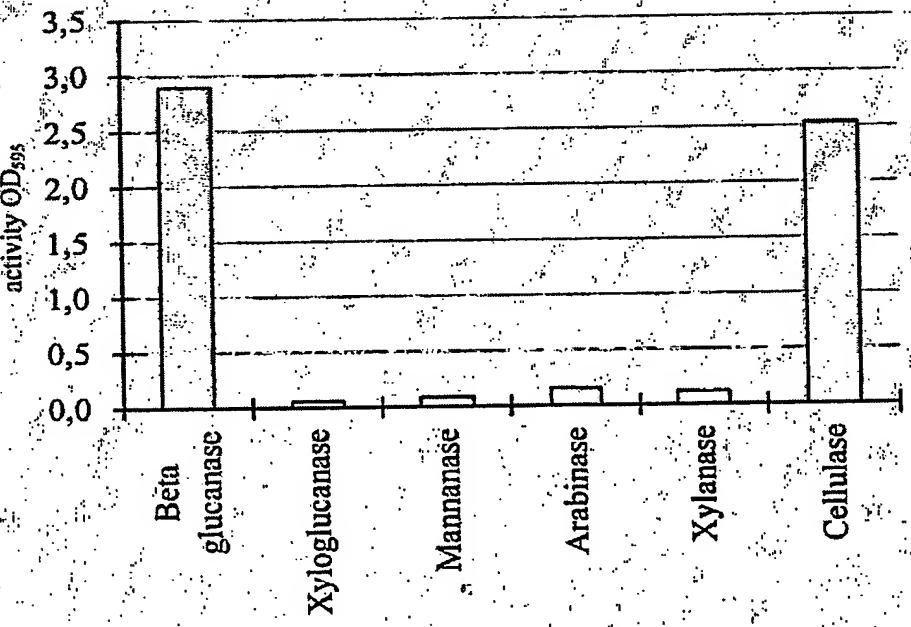

Fig. 7

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Fig. 8

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Fig. 9

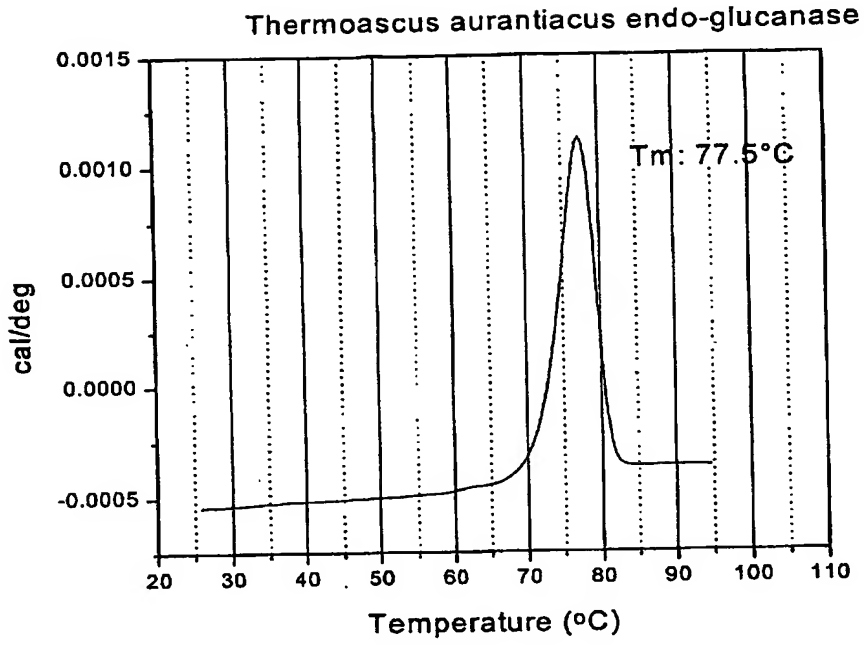
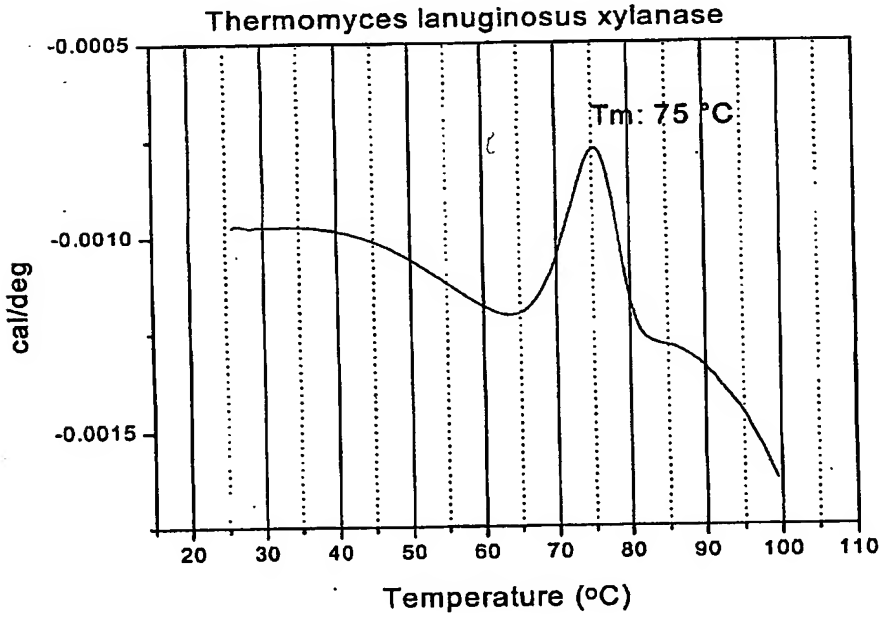



Fig. 10



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# 中国微生物菌种保藏管理委员会
# 普通微生物中心
# China General Microbiological Culture Collection Center (CGMCC)

Address: Institute of Microbiology, Chinese Academy of Sciences, P. O. Box 2714, Beijing 100080, P. R. China
Telephone: 86-10-62555614 Fax: 86-10-62542758 E-mail: cgmcc@sun.im.ac.cn

## 受理通知书
## NOTIFICATION OF RECEIPT

Deposition of culture for patent purposes
under Budapest Treaty

CGMCC NO. 0670

1. Name and address of the depositor or agent

吴文平 Wu Wen Ping
Novozymes (China) Investment Co. Ltd
22 Xinxi Zhong Lu, Shangdi Zone, Haidian District, Beijing 100080, P. R. China

2. Strain reference given by depositor

T002-5

3. Deposited microorganisms appended

☐ Scientific description

☒ Proposed taxonomic name

*Thermoascus aurantiacus*

4. The deposited microorganism has been received and numbered as CGMCC No. 0670 on Dec. 27, 2001 .

Your application for converting original deposit to Budapest Treaty deposit has been received on Dec. 27, 2001 .

Signature of Head of CGMCC Zhou Yuguang

Date Dec. 27, 2001

中国微生物菌种保藏管理委员会
普通微生物中心
China General Microbiological Culture Collection Center(CGMCC)

Address: Institute of Microbiology, Chinese Academy of Sciences, P. O. Box 2714, Beijing 100080, P. R. China
Telephone: 86-10-62555614 Fax: 86-10-62560912 E-mail: CGMCC@sun.im.ac.cn

# 存活性报告书
# VIABILITY STATEMENT

Deposition of culture for patent purposes
under Budapest Treaty

**CGMCC NO. 0670**

1. Name and address of the depositor or agent

吴文平 Wu Wen Ping
Novozymes (China) Investment Co. Ltd
22 Xinxi Zhong Lu, Shangdi Zone, Haidian District, Beijing 100080, P. R. China

2. Strain reference given by depositor

T002-5

3. The deposited microorganism has been received and numbered as CGMCC No. 0670 on Dec. 27, 2001 .
The viability test has already been performed on Dec. 30, 2001. The result is

☒ viable; ☐ no longer viable

4. The conditions under which the viability test has been performed

Medium: PDA, pH6.5
Temperature: 45℃

Signature of Head of CGMCC Zhou Yuguang

Date Dec. 30, 2001

BUDAPEST TREATY ON THE INTERNATIONAL RECOGNITION OF THE DEPOSIT OF MICROORGANISMS FOR THE PURPOSES OF PATENT PROCEDURE

2001-15947-01

INTERNATIONAL FORM

Modtaget PVS

25 JAN. 2002

Novozymes A/S
Krogshøjvej 36
2880 Bagsvaerd
Denmark

RECEIPT IN THE CASE OF AN ORIGINAL DEPOSIT
issued pursuant to Rule 7.1 by the
INTERNATIONAL DEPOSITARY AUTHORITY
identified at the bottom of this page

| I. IDENTIFICATION OF THE MICROORGANISM | |
|---|---|
| Identification reference given by the DEPOSITOR:<br>NN049586 | Accession number given by the INTERNATIONAL DEPOSITARY AUTHORITY:<br>DSM 14541 |

II. SCIENTIFIC DESCRIPTION AND/OR PROPOSED TAXONOMIC DESIGNATION

The microorganism identified under I. above was accompanied by:

( ) a scientific description
(X) a proposed taxonomic designation

(Mark with a cross where applicable).

III. RECEIPT AND ACCEPTANCE

This International Depositary Authority accepts the microorganism identified under I. above, which was received by it on 2001-09-28 (Date of the original deposit)[1].

IV. RECEIPT OF REQUEST FOR CONVERSION

The microorganism identified under I above was received by this International Depositary Authority on (date of original deposit) and a request to convert the original deposit to a deposit under the Budapest Treaty was received by it on (date of receipt of request for conversion).

V. INTERNATIONAL DEPOSITARY AUTHORITY

| | |
|---|---|
| Name: DSMZ-DEUTSCHE SAMMLUNG VON MIKROORGANISMEN UND ZELLKULTUREN GmbH<br>Address: Mascheroder Weg 1b<br>D-38124 Braunschweig | Signature(s) of person(s) having the power to represent the International Depositary Authority or of authorized official(s):<br>Dagmar Tilker<br>Date: 2001-10-01 |

[1] Where Rule 6.4 (d) applies, such date is the date on which the status of international depositary authority was acquired.

Form DSMZ-BP/4 (sole page) 0196

BUDAPEST TREATY ON THE INTERNATIONAL RECOGNITION OF THE DEPOSIT OF MICROORGANISMS FOR THE PURPOSES OF PATENT PROCEDURE

2001-15947-01

INTERNATIONAL FORM

Modtaget PVS
25 JAN. 2002

Novozymes A/S
Krogshøjvej 36
2880 Bagsvaerd
Denmark

VIABILITY STATEMENT
issued pursuant to Rule 10.2 by the
INTERNATIONAL DEPOSITARY AUTHORITY
identified at the bottom of this page

| I. DEPOSITOR | II. IDENTIFICATION OF THE MICROORGANISM |
|---|---|
| Name: Novozymes A/S<br>Krogshøjvej 36<br>Address: 2880 Bagsvaerd<br>Denmark | Accession number given by the INTERNATIONAL DEPOSITARY AUTHORITY:<br>DSM 14541<br>Date of the deposit or the transfer[1]:<br>2001-09-28 |

III. VIABILITY STATEMENT

The viability of the microorganism identified under II above was tested on 2001-09-28 [2].
On that date, the said microorganism was

(X)[3] viable

( )[3] no longer viable

IV. CONDITIONS UNDER WHICH THE VIABILITY TEST HAS BEEN PERFORMED[4]

V. INTERNATIONAL DEPOSITARY AUTHORITY

| | |
|---|---|
| Name: DSMZ-DEUTSCHE SAMMLUNG VON MIKROORGANISMEN UND ZELLKULTUREN GmbH<br>Address: Mascheroder Weg 1b<br>D-38124 Braunschweig | Signature(s) of person(s) having the power to represent the International Depositary Authority or of authorized official(s):<br>Date: 2001-10-01 |

[1] Indicate the date of original deposit or, where a new deposit or a transfer has been made, the most recent relevant date (date of the new deposit or date of the transfer).
[2] In the cases referred to in Rule 10.2(a) (ii) and (iii), refer to the most recent viability test.
[3] Mark with a cross the applicable box.
[4] Fill in if the information has been requested and if the results of the test were negative.

Form DSMZ-BP/9 (sole page) 0196